

MAY 20 2003

-1-

(Translation)

Mailed: April 15, 2003

NOTIFICATION OF REASONS FOR REJECTION

RECEIVED

Patent Application No.: 2000-247729

MAY 22 2003

Examiner's Notice Date: April 8, 2003

TECH CENTER 1600/2900

Examiner: Keiko Nagai

This application is rejected on the grounds stated below. Any opinion about the rejection must be filed within 60 DAYS of the mailing date hereof.

REASON

The invention is unpatentable under Section 29 (2) of the Patent Law, as being such that the invention could easily have been made by a person with ordinary skill in the art to which the invention pertains, on the basis of the invention described in the following publication(s) distributed in Japan or a foreign country prior to this application.

REMARKS

(1) Claim 1 : Reference 1

Reference 1 discloses a nucleotide sequence of an estrogen receptor gene of medaka fish. Here, it is easily achievable for a person having ordinary skill in the art to prepare a probe based on the nucleotide sequence of the gene discussed in Reference 1, thus obtaining polynucleotide including upstream and downstream regions of the estrogen receptor gene of medaka fish and determining its nucleotide sequence.

(2) Claim 2 : Reference 1

Reference 1 discloses a nucleotide sequence of an estrogen receptor gene of medaka fish. Here, it is easily achievable for a person having ordinary

discussed in Reference 1, thus obtaining an estrogen receptor gene of medaka fish and determining its nucleotide sequence.

(3) Claim 3 : Reference 1

Reference 1 discloses a nucleotide sequence of an estrogen receptor gene of medaka fish, and an amino acid sequence of the estrogen receptor. Here, it is easily achievable for a person having ordinary skill in the art to prepare a probe based on the nucleotide sequence of the gene discussed in Reference 1, thus obtaining an estrogen receptor gene of medaka fish and determining its nucleotide sequence and an amino acid sequence encoded by it.

(4) Claim 4 : Reference 1

As described in remarks (1) and (2) above, it is easily achievable for a person having ordinary skill in the art to obtain polynucleotide recited in Claims 1 and 2. Assembly of a recombinant vector is merely a well-known means.

(5) Claim 5 : Reference 1

Introduction of a gene to a host in order to examine an in-vivo function of a product of an isolated gene is well-known means.

The claims not mentioned in this Official Action are not rejected. If a new reason for rejection is noticed, a further Official Action will be issued.

Reference Cited:

1. Jpn. Pat. Appln. KOKAI Publication 2000-201688

Prior Art Search Report

Searched Field: IPC 7th ed. C12N 15/00

SwissProt/PIR/GeneSeq

Genbank/EMBL/DDBJ/GeneSeq

BIOSIS

MEDLINE

WPIDS

Prior-Art Document(s):

Winn R., Marine Environmental Research, vol. 46 (1-5), p.130 (1998)

pp. 192-199 (1994)

Gray M.A. et al., Environmental Toxicology and Chemistry, vol. 18(11), pp. 2587-2594 (1999)

The result of this prior art search does not constitute the reasons for rejection.

Mailing Date: April 15, 2005

整理番号 A 0 0 0 0 0 3 8 8 5

発送番号 1 2 1 2 6 4
発送日 平成 1 5 年 4 月 1 5 日

1 / 3

拒絶理由通知書

特許出願の番号 特願 2 0 0 0 - 2 4 7 7 2 9
起案日 平成 1 5 年 4 月 8 日
特許庁審査官 長井 啓子 9 1 2 3 4 N 0 0
特許出願人代理人 鈴江 武彦 (外 5 名) 様
適用条文 第 2 9 条第 2 項

15.6.14

この出願は、次の理由によって拒絶をすべきものである。これについて意見があれば、この通知書の発送の日から 6 0 日以内に意見書を提出して下さい。

理 由

この出願の下記の請求項に係る発明は、その出願前日本国内又は外国において頒布された下記 of 刊行物に記載された発明に基いて、その出願前にその発明の属する技術の分野における通常の知識を有する者が容易に発明をすることができたものであるから、特許法第 2 9 条第 2 項の規定により特許を受けることができない。

記 (引用文献等については引用文献等一覧参照)

(1) 請求項 1 : 引用文献 1

引用文献 1 には、メダカのエストロゲンレセプター遺伝子の塩基配列が開示されている。引用文献 1 記載の遺伝子の塩基配列を基にしてプローブを作成して、メダカのエストロゲンレセプター遺伝子の上流及び下流の領域を含むポリヌクレオチドを得てその塩基配列を決定することは、当業者が容易になし得る程度のことにすぎない。

(2) 請求項 2 : 引用文献 1

引用文献 1 には、メダカのエストロゲンレセプター遺伝子の塩基配列が開示されている。引用文献 1 記載の遺伝子の塩基配列を基にしてプローブを作成して、メダカのエストロゲンレセプター遺伝子を得てその塩基配列を決定することは、当業者が容易になし得る程度のことにすぎない。

(3) 請求項 3 : 引用文献 1

引用文献 1 には、メダカのエストロゲンレセプター遺伝子の塩基酸配列及び当該エストロゲンレセプターのアミノ酸配列が開示されている。引用文献 1 記載の

広島大学長

遺伝子の塩基配列を基にしてプローブを作成して、メダカのエストロゲンレセプター遺伝子を得てその塩基配列及びそれがコードするアミノ酸配列を決定することとは、当業者が容易になし得る程度のことにはすぎない。

(4) 請求項4：引用文献1

請求項1及び請求項2記載のポリヌクレオチドを得ることが、引用文献1の記載に基づいて当業者が容易になし得たことは、上記(1)及び(2)で説明したとおりである。組み換えベクターを構築することは常套手段にすぎない。

(5) 請求項5：引用文献1

単離した遺伝子の産物の生体内機能を探る等の目的で、宿主に遺伝子導入することは、常套手段である。

この拒絶理由通知書中で指摘した請求項以外の請求項に係る発明については、現時点では、拒絶の理由を発見しない。拒絶の理由が新たに発見された場合には拒絶の理由が通知される。

引用文献等一覧

1. 特開2000-201688号公報

この拒絶理由通知書に不明な点がある場合、または、この案件について面接を希望する場合は、

特許審査第三部生命工学 長井 啓子

Tel. 03-3581-1101 (特許庁代表)

Fax. 03-3501-0491

までご連絡下さい。

先行技術文献調査結果の記録

・調査した分野 IPC第7版 C12N 15/00
SwissProt/PIR/GeneSeq
Genbank/EMBL/DDBJ/GeneSeq

MEDELIAL

WPIDS

ID AAA92174 standard; DNA; 1728 BP.
 XX
 AC AAA92174; **XP-002181484**
 XX

DT 05-JAN-2001 (first entry)
 XX

DE Oryzias lapites oestrogen receptor encoding DNA SEQ ID NO:2.
 XX

KW Oryzias lapites; oestrogen receptor; ds.
 XX

OS Oryzias lapites.
 XX

PN JP2000201688-A.
 XX

PD 25-JUL-2000.
 XX

PF 06-APR-1999; 99JP-0098787.
 XX

PR 10-NOV-1998; 98JP-0319465.
 XX

PA (SUMO) SUMITOMO CHEM CO LTD.
 XX

DR WPI; 2000-567950/53.
 DR

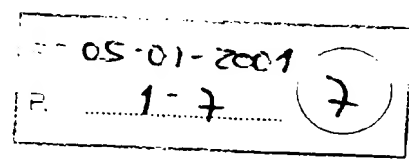
DR P-PSDB; AAB20897.
 XX

PT An estrogen receptor gene and its application -
 XX

PS Claim 3; Page 11-13; 23pp; Japanese.
 XX

CC The present sequence encodes an oestrogen receptor derived from
 CC Oryzias lapites. Also described are: (1) a vector comprising the
 CC oestrogen receptor gene; (2) a transformant prepared by introducing
 CC the oestrogen receptor gene or vector from (1) into a host cell;
 CC (3) a method for the preparation of an oestrogen receptor comprising
 CC culturing the transformant from (2) to produce the oestrogen receptor;
 CC and (4) a method for the evaluation of oestrogen receptor-activating
 CC ability of a chemical substance in which the chemical substance is
 CC reacted with a transformant prepared by introducing a reporter gene
 CC connected downstream of a transcription controlling region containing
 CC an oestrogen response sequence and the above oestrogen receptor gene to
 CC an oestrogen-nonendogenous host cell. The transformant can be used for
 CC the evaluation of oestrogen receptor-activating ability of a chemical
 CC substance.

XX
 SQ Sequence 1728 BP; 378 A; 514 C; 497 G; 339 T; 0 other;
 atgtaccctg aagagagccg ggggttctgga ggggtggtctg ctgtggacct tttggaaggg 60
 acgtacgact atgccgcccc caaccctgcc acgactcccc ttacagcca gtccagcacc 120
 ggctactact ctgctcccc ggaacaaac ggacccccct cagaaggcag tctgcagtcc 180
 ctgggcagtg ggccgacgag cctctgtgtg tttgtgccct ccagccccag actcagtccc 240
 tttatgcatc caccagcca ccactatctg gaaaccatt ccacgccgt ttacagatcc 300
 agccaccagg gagcctccag ggaggaccag tgcggctccc gggaggacac gtgcagcctg 360
 ggggagttag gcgcccggagc cggggctggg gggtttgaga tggccaaaga cacgcgtttc 420
 tgcgccgtgt gcagcgacta cgcctctggg taccactatg ggtgtgtgtc ttgtgagggc 480
 tgcaaggcct tcttcaagag gagcatccag ggtcacaatg actatatgtg ccagcgacc 540
 aatcagtga ctattgacag aaatcgaagg aagggtgtgc aggtttgtcg tcttaggaag 600
 tgttacgaag tgggaatgat gaaaggcgtg gtgcgcaagg accgcattcg cattttacg 660
 cgtgacaaac ggcggacagg cgttggtgat ggagacaagg ttgtaaaggg tcaggagcat 720
 aaaacggtgc attatgatg aaggaaacgc agcagcacag gaggaggagg aggaggagg 780
 ggaggaagac tgtctgtgac cagcatacct cctgagcagg tgctgtcct ccttcagggc 840
 gccgagcccc cgatactctg ctgcgctcag aagttgagcc gaccgtacac cgaggtcacc 900
 atgatgacct tgctcaccag catggcagac aaggagctgg tccacatgat cgcctgggcc 960
 aagaagctcc cagggtttct gcagctgtcc ctgcacgac aggtgtgtgt gctggagagc 1020
 tcgtggctgg aggtgtcat gatcgccctc atttgagggt ccatccactg tcccgggaag 1080
 ctcattcttg cacaagacct catcctggac aggaatgagg gagactgcgt ggaaggcatg 1140



acggagatct	togacatgct	gctggccact	gcttcccgt	tccgtgtgct	caaactcaaa	1200
cctgaggaat	tcgtctgct	caaagctatt	attttactca	actccggtgc	tttttctttc	1260
tgacccggca	ccatggagcc	acttcacaac	agcgcggcgg	ttcagagcat	gctggacacc	1320
atcacagacg	cactcattca	ttacatcagt	cagtcgggtt	acttgcccca	ggagcaggcg	1380
agacggcagg	cccagccgct	cctgctgctc	tcccacatca	ggcacatgag	caacaaaggc	1440
atggagcacc	tctacagcat	gaagtgcagg	aacaaagtcc	ctctttatga	cctcctactg	1500
gagatgctcg	atgcccaccg	cctgcaccac	cccgtcagag	ccccccagtc	cttgtcccaa	1560
gtcgacagag	accctccctc	caccagcagc	ggcgggggtg	gaatcgctcc	cggttctata	1620
tcagcatctc	gaggcagaat	cgagagtcgg	agcagaggcc	cctttgctcc	cagtgtcctt	1680
cagtatggag	ggtcgcgtcc	tgactgcacc	ccggcccttc	aagactga		1728

//

>>GSN:AAA92174 Oryzias lapites oestrogen recept (1728 nt)
initn: 8586 initl: 8586 opt: 8586 Z-score: 8271.0 bits: 1544.2 E(): 0
99.653% identity (99.653% ungapped) in 1728 nt overlap (211-1938:1-1728)

```

      190      200      210      220      230      240
EP0111 CGCCTCTCGCCCCGTGACCCCTCGGTGACATGTACCCCTGAAGAGAGCCGGGGTTCTGGA
      :
GSN:AA      ATGTACCCTGAAGAGAGCCGGGGTTCTGGA
                        10      20      30

      250      260      270      280      290      300
EP0111 GGGGTGGCTGCTGTGGACTTTTGGGAAGGGACGTACGACTATGCCGCCCCCAACCCCTGCC
      :
GSN:AA GGGGTGGCTGCTGTGGACCTTTTGGGAAGGGACGTACGACTATGCCGCCCCCAACCCCTGCC
      40      50      60      70      80      90

      310      320      330      340      350      360
EP0111 ACGACTCCCCCTTACAGCCAGTCCAGCACCGGCTACTACTCTGCTCCCTGGAAACAAAC
      :
GSN:AA ACGACTCCCCCTTACAGCCAGTCCAGCACCGGCTACTACTCTGCTCCCTGGAAACAAAC
      100      110      120      130      140      150

      370      380      390      400      410      420
EP0111 GGACCCCCCTCAGAAGGCAGTCTGCAGTCCCTGGGCAGTGGGCCGACGAGCCCTCTGGTG
      :
GSN:AA GGACCCCCCTCAGAAGGCAGTCTGCAGTCCCTGGGCAGTGGGCCGACGAGCCCTCTGGTG
      160      170      180      190      200      210

      430      440      450      460      470      480
EP0111 TTTGTGCCCTCCAGCCCCAGACTCAGTCCCTTTATGCATCCACCCAGCCCACTATCTG
      :
GSN:AA TTTGTGCCCTCCAGCCCCAGACTCAGTCCCTTTATGCATCCACCCAGCCCACTATCTG
      220      230      240      250      260      270

      490      500      510      520      530      540
EP0111 GAAACCACTTCCACGCCCGTTTACAGATCCAGCCACCAGGGAGCCTCCAGGGAGGACCAG
      :
GSN:AA GAAACCACTTCCACGCCCGTTTACAGATCCAGCCACCAGGGAGCCTCCAGGGAGGACCAG
      280      290      300      310      320      330

      550      560      570      580      590      600
EP0111 TGCGGCTCCCGGGAGGACACGTGCAGCCTGGGGGAGTTAGGCGCCGGAGCCGGGGCTGGG
      :
GSN:AA TGCGGCTCCCGGGAGGACACGTGCAGCCTGGGGGAGTTAGGCGCCGGAGCCGGGGCTGGG
      340      350      360      370      380      390

      610      620      630      640      650      660
EP0111 GGGTTTGAGATGGCCAAAGACACGCGTTTCTGCGCCGTGTGCAGCGACTACGCCTCTGGG
      :
GSN:AA GGGTTTGAGATGGCCAAAGACACGCGTTTCTGCGCCGTGTGCAGCGACTACGCCTCTGGG
      400      410      420      430      440      450

      670      680      690      700      710      720
EP0111 TACCACTATGGGGTGTGGTCTTGTGAGGGCTGCAAGGCCTTCTTCAAGAGGAGCATCCAG
      :
GSN:AA TACCACTATGGGGTGTGGTCTTGTGAGGGCTGCAAGGCCTTCTTCAAGAGGAGCATCCAG
      460      470      480      490      500      510

      730      740      750      760      770      780
EP0111 GGTCACAATGACTATATGTGCCCAGCGACCAATCAGTGCACTATTGACAGAAATCGGAGG
      :
GSN:AA GGTCACAATGACTATATGTGCCCAGCGACCAATCAGTGCACTATTGACAGAAATCGAAGG
      520      530      540      550      560      570
```


UNSCDOCID: YD 2181494A 1

1450 1460 1470 1480 1490 1500

EP0111 ATTTTACTCAACTCCGGTGCTTTTTCTTTCTGACCGGCACCATGCGAGCCACTTCACAAC
:
GSN:AA ATTTTACTCAACTCCGGTGCTTTTTCTTTCTGACCGGCACCATGCGAGCCACTTCACAAC
1240 1250 1260 1270 1280 1290

1510 1520 1530 1540 1550 1560

EP0111 AGCGCGGCGGTTCAGAGCATGCTGGACACCATCACAGACGGAATCATTACATCAGT
:
GSN:AA AGCGCGGCGGTTCAGAGCATGCTGGACACCATCACAGACGGAATCATTACATCAGT
1300 1310 1320 1330 1340 1350

1570 1580 1590 1600 1610 1620

EP0111 CAGTCGGGTTACTTTGGCCCAGGAGCAGGCGAGACGGCAGGCCAGCTGCTCCTGCTGCTC
:
GSN:AA CAGTCGGGTTACTTTGGCCCAGGAGCAGGCGAGACGGCAGGCCAGCTGCTCCTGCTGCTC
1360 1370 1380 1390 1400 1410

1630 1640 1650 1660 1670 1680

EP0111 TCCCACATCAGGCACATGAGCAAACAAGGCATGAGACCTCTACAGCATGAAGTCCAAG
:
GSN:AA TCCCACATCAGGCACATGAGCAAACAAGGCATGAGACCTCTACAGCATGAAGTCCAAG
1420 1430 1440 1450 1460 1470

1690 1700 1710 1720 1730 1740

EP0111 AACAAAGTCCCTCTTTATGACCTCCTACTGAGATGCTCGATGGSCACCGCCTGCACCA
:
GSN:AA AACAAAGTCCCTCTTTATGACCTCCTACTGAGATGCTCGATGGSCACCGCCTGCACCA
1480 1490 1500 1510 1520 1530

1750 1760 1770 1780 1790 1800

EP0111 CCCGTCAGAGCACCCCAGTCCTTGTCCCAAGTCGACAGAGACCTCCCTCCACCAGCAGC
:
GSN:AA CCCGTCAGAGCCCCCAGTCCTTGTCCCAAGTCGACAGAGACCTCCCTCCACCAGCAGC
1540 1550 1560 1570 1580 1590

1810 1820 1830 1840 1850 1860

EP0111 GGCGGGGGTGGAATCGCTCCCGGTTCTATATCAGCATCTCGAGGCAGAATCGAGAGTCCG
:
GSN:AA GGCGGGGGTGGAATCGCTCCCGGTTCTATATCAGCATCTCGAGGCAGAATCGAGAGTCCG
1600 1610 1620 1630 1640 1650

1870 1880 1890 1900 1910 1920

EP0111 AGCAGAGGCCCTTTGCTCCCAGTGTCTTCAGTATGGAGGGTCGCGTCCTGACTGCACC
:
GSN:AA AGCAGAGGCCCTTTGCTCCCAGTGTCTTCAGTATGGAGGGTCGCGTCCTGACTGCACC
1660 1670 1680 1690 1700 1710

1930 1940 1950 1960 1970 1980

EP0111 CCGGCCCTTCAAGACTGAGCACACAGTCCAAGGCCCTTTTTTGTGGCTCAAGGGTTCAG
:
GSN:AA CCGGCCCTTCAAGACTGA
1720

17
ID AAB20897 standard; Protein; 575 AA.

XX
AC AAB20897;

XX
DT 05-JAN-2001 (first entry)

XX
DE Oryzias lapites oestrogen receptor protein SEQ ID NO:1.

XX
KW Oryzias lapites; oestrogen receptor.

XX
OS Oryzias lapites.

XX
PN JP2000201688-A.

XX
PD 25-JUL-2000.

XX
PF 06-APR-1999; 99JP-0098787.

XX
PR 10-NOV-1998; 98JP-0319465.

XX
PA (SUMO) SUMITOMO CHEM CO LTD.

XX
DR WPI; 2000-567950/53.

XX
DR N-PSDB; AAA92174.

XX
PT An estrogen receptor gene and its application

XX
PS Claim 1; Page 9-10; 23pp; Japanese.

XX
CC The present sequence represents an oestrogen receptor derived from
CC Oryzias lapites. Also described are: (1) a vector comprising the
CC oestrogen receptor gene; (2) a transformant prepared by introducing
CC the oestrogen receptor gene or vector from (1) into a host cell;
CC (3) a method for the preparation of an oestrogen receptor comprising
CC culturing the transformant from (2) to produce the oestrogen receptor;
CC and (4) a method for the evaluation of oestrogen receptor-activating
CC ability of a chemical substance in which the chemical substance is
CC reacted with a transformant prepared by introducing a reporter gene
CC connected downstream of a transcription controlling region containing
CC an oestrogen response sequence and the above oestrogen receptor gene to
CC an oestrogen-nonendogenous host cell. The transformant can be used for
CC the evaluation of oestrogen receptor-activating ability of a chemical
CC substance.

XX
SQ Sequence. 575 AA;

SQ 37 A; 38 R; 10 N; 27 D; 0 B; 18 C; 24 Q; 29 E; 0 Z; 58 G; 19 H;
SQ 22 I; 61 L; 24 K; 19 M; 14 F; 37 P; 58 S; 29 T; 4 W; 20 Y; 27 V;
SQ 0 Others;

mypeesrgsg gvaavdlleg tydyaapnpa ttplysqsst gyysapletn gppsegsllqs
lgsqptsplv fvpssprlsp fmhppsghyl ettstpvrys shqgasredq cgsredtcs1
gelgagagag gfemakdtrf cavcsdyasg yhygvwsceg ckaffkrsiq ghndymcpat
nqctidrnrr kgcqacrlrk cyevgmkkqg vrkdririlr rdkrrtgvd qdkvvkqgeh
ktvhydgrkr sstggggggg ggrlsvtsip peqvlllllg aeppilcsrq klsrpytevt
mntlltsmad kelvhmiawa kklpgflqls lhdqvlles swlevlmigl iwrsihcpqk
lifagdlild negdcvegm teifdmllat asrfrvlklk peefvcl kai illnsgafsf
ctgtmeplhn saavqsmltd itdalihiyis qsgylaqeqa rrqaqpllll shirhmsnkq
mehlysmkck nkvpdydlld emldahrlhb pvrpqslsq vdrdpstss ggggiapgsi
sasrgriesp srgpfapsvl qyggsrpdct palqd

//

>>GSP:AAB20897 Oryzias lapites oestrogen recept (575 aa)
 initn: 3905 initl: 3905 opt: 3905 Z-score: 2879.0 bits: 544.9 E(): 2.1e-152
 Smith-Waterman score: 3905; 99.478% identity (99.478% ungapped) in 575 aa overlap
 (210-1934:1-575)

```

      210      240      270      300      330      360
EP0111 MYPEESRSGSGGVAADVFLLEGTYDYAAPNPATTPLYSQSSTGYYSAPLETNGPPSEGLQSLQ
      :
GSP:AA MYPEESRSGSGGVAADVFLLEGTYDYAAPNPATTPLYSQSSTGYYSAPLETNGPPSEGLQSLQ
      10      20      30      40      50      60

```

```

      390      420      450      480      510      540
EP0111 LGSGPTSPLVFPSSPRLSPFMHPPSHHYLETTSTPVYRSSHQASREDQCGSREDTCSL
      :
GSP:AA LGSGPTSPLVFPSSPRLSPFMHPPSHHYLETTSTPVYRSSHQASREDQCGSREDTCSL
      70      80      90      100      110      120

```

```

      570      600      630      660      690      720
EP0111 GELGAGAGAGGFEMAKDTRFCVCSYASGYHYGVWSCEGCKAFFKRSIQGHNDYMCPAT
      :
GSP:AA GELGAGAGAGGFEMAKDTRFCVCSYASGYHYGVWSCEGCKAFFKRSIQGHNDYMCPAT
      130      140      150      160      170      180

```

```

      750      780      810      840      870      900
EP0111 NQCTIDRNRRKSCQACRLRKCYEVGMMKGGVVRKDRIRILRRDKRRTGVGDGDKVVKGQEH
      :
GSP:AA NQCTIDRNRRKSCQACRLRKCYEVGMMKGGVVRKDRIRILRRDKRRTGVGDGDKVVKGQEH
      190      200      210      220      230      240

```

```

      930      960      990      1020      1050      1080
EP0111 KTVHYDGRKRSSTGGGGGGGGGRLSVTSIPPEQVLLLLLQGAEPPIILCSRQKLSRPYTEVT
      :
GSP:AA KTVHYDGRKRSSTGGGGGGGGGRLSVTSIPPEQVLLLLLQGAEPPIILCSRQKLSRPYTEVT
      250      260      270      280      290      300

```

```

      1110      1140      1170      1200      1230      1260
EP0111 MMTLLTSMADKELVHMIWAKKLPGLQLSLHDQVLLLESSWLEVLMIGLIWRSIHCPGK
      :
GSP:AA MMTLLTSMADKELVHMIWAKKLPGLQLSLHDQVLLLESSWLEVLMIGLIWRSIHCPGK
      310      320      330      340      350      360

```

```

      1290      1320      1350      1380      1410      1440
EP0111 LIFAQDLILDRNEGDCVEGMTEIFDMLLATASRFRVLKLPKEEFVCLKAIILLNSGAFSF
      :
GSP:AA LIFAQDLILDRNEGDCVEGMTEIFDMLLATASRFRVLKLPKEEFVCLKAIILLNSGAFSF
      370      380      390      400      410      420

```

```

      1470      1500      1530      1560      1590      1620
EP0111 CTGTMEPLHNSAAVQSMMLDTITDALIHYSQSGYLAQEQAARRQAQLLLLLSHIRHMSNKG
      :
GSP:AA CTGTMEPLHNSAAVQSMMLDTITDALIHYSQSGYLAQEQAARRQAQLLLLLSHIRHMSNKG
      430      440      450      460      470      480

```

```

      1650      1680      1710      1740      1770      1800
EP0111 MEHLYSMKCKNKVPLYDLLLEMLDAHRLHHPVRAPQSLSQVDRDPPSTSSGGGGIAPGSI
      :
GSP:AA MEHLYSMKCKNKVPLYDLLLEMLDAHRLHHPVRAPQSLSQVDRDPPSTSSGGGGIAPGSI
      490      500      510      520      530      540

```

```

      1830      1860      1890      1920
EP0111 SASRGRIESPSRGPFAPSVLQYGGSRPDCTPALQD
      :
GSP:AA SASRGRIESPSRGPFAPSVLQYGGSRPDCTPALQD
      550      560      570

```

ID AAA92175 standard; DNA; 1863 BP.

XX

AC AAA92175;

XP-002181483

XX

DT 05-JAN-2001 (first entry)

XX

DE Oryzias lapites oestrogen receptor encoding DNA-SEQ-ID NO:4.

XX

KW Oryzias lapites; oestrogen receptor, ds.

XX

OS Oryzias lapites.

XX

PN JP2000201688-A.

XX

PD 25-JUL-2000.

XX

PF 06-APR-1999; 99JP-0098787.

XX

PR 10-NOV-1998; 98JP-0319465.

XX

PA (SUMO) SUMITOMO CHEM CO LTD.

XX

DR WPI; 2000-567950/53.

DR

P-PSDB; AAB20898.

XX

PT An estrogen receptor gene and its application

XX

PS Claim 4; Page 15-17; 23pp; Japanese.

XX

CC The present sequence encodes an oestrogen receptor derived from
CC Oryzias lapites. Also described are: (1) a vector comprising the
CC oestrogen receptor gene; (2) a transformant prepared by introducing
CC the oestrogen receptor gene or vector from (1) into a host cell;
CC (3) a method for the preparation of an oestrogen receptor comprising
CC culturing the transformant from (2) to produce the oestrogen receptor;
CC and (4) a method for the evaluation of oestrogen receptor-activating
CC ability of a chemical substance in which the chemical substance is
CC reacted with a transformant prepared by introducing a reporter gene
CC connected downstream of a transcription controlling region containing
CC an oestrogen response sequence and the above oestrogen receptor gene to
CC an oestrogen-nonendogenous host cell. The transformant can be used for
CC the evaluation of oestrogen receptor-activating ability of a chemical
CC substance.

XX

SQ Sequence 1863 BP; 406 A; 565 C; 531 G; 361 T; 0 other;

atgagtaaga	gacagagctc	ggtgcagatc	aggcagctgt	tcggaccagc	actcagatcc	60
aggatcagcc	cagcctcctc	agagctggag	accctctccc	cacctcgctt	ctcgccccgt	120
gacccccctc	gtagacatga	ccctgaagag	agccgggggt	ctggaggggt	ggctgctgtg	180
gaccttttgg	aagggacgta	cgactatgcc	gcccccaacc	ctgccacgac	tcccccttac	240
agccagtgca	gcaccggcta	ctactctgct	cccctggaaa	caaacggacc	cccctcagaa	300
ggcagtcctg	agtcctctgg	cagtgggccc	acgagccctc	tggtgtttgt	gcccctccagc	360
cccagactca	gtccctttat	gcatccaccc	agccaccact	atctggaaac	caattccacg	420
cccgtttaca	gatccagcca	ccagggagcc	tccagggagg	accagtgcgg	ctcccggggg	480
gacacgtgca	gcctggggga	gttaggcgcc	ggagccgggg	ctgggggggt	tgagatggcc	540
aaagacacgc	gtttctgcgc	cgtgtgcagc	gactacgcct	ctgggtacca	ctatgggggtg	600
tggtcttggt	agggctgcaa	ggccttcttc	aagaggagca	tccagggtca	caatgactat	660
atgtgcccag	cgaccaatca	gtgcactatt	gacagaaatc	gaaggaaggg	ctgtcaggct	720
tgctgctcta	ggaagtgtta	cgaagtggga	atgatgaaag	gcgggtgtgc	caaggaccgc	780
attcgcatth	tacggcgtga	caaacggcgg	acaggcggtg	gtgatggaga	caaggttgta	840
aagggtcagg	agcataaaac	ggtgcattat	gatggaagga	aacgcagcag	cacaggagga	900
ggaggaggag	gaggaggagg	aagactgtct	gtgaccagca	tacctctga	gcagggtgctg	960
ctcctccttc	agggcgccga	gcccccgata	ctctgctcgc	gtcagaagtt	gagccgaccg	1020
tacaccgagg	tcaccatgat	gacctgtctc	accagcatgg	cagacaagga	gctgggtccac	1080
atgatcgctt	gggccaagaa	gctcccaggt	tttctgcagc	tgctccctga	cgatcagggtg	1140

ctgctgctgg	agagctcgtg	gctggaggtg	ctcatgatcg	gcctcatttg	gaggtccatc	1200
cactgtcccg	ggaagctcat	ctttgcacaa	gacctcatcc	tggacaggaa	tgagggagac	1260
tgcgtggaag	gcatgacgga	gatcttcgac	atgctgctgg	ccactgcttc	ccgcttcogt	1320
gtgctcaaac	tcaaacctga	ggaattcgtc	tgctcaaag	ctattatatt	actcaactcc	1380
ggtgcttttt	ctttctgcac	cggcaccatg	gagccacttc	acaacagcgc	ggcggttcag	1440
agcatgctgg	acaccatcac	agacgcactc	attcattaca	tcagtcagtc	gggttacttg	1500
gcccaggagc	aggcgagacg	gcaggcccag	ccgctcctgc	tgctctccca	catcaggcac	1560
atgagcaaca	aaggcatgga	gcacctctac	agcatgaagt	gcaagaacaa	agtccctctt	1620
tatgacctcc	tactggagat	gctcgatgcc	caccgcctgc	accacccogt	cagagccccc	1680
cagtccttgt	cccaagtcga	cagagaccct	ccctccacca	gcagcggcgg	gggtggaatc	1740
gctcccgggt	ctatatcagc	atctcgaggc	agaatcgaga	gtccgagcag	aggccctttt	1800
gctcccagtg	tccttcagta	tggaggggtc	cgtcctgact	gcaccccggc	ccttcaagac	1860
tga						1863

//

>>GSN:AAA92175 Oryzias lapites oestrogen recept (1863 nt)
initn: 9261 initl: 9261 opt: 9261 Z-score: 8922.0 bits: 1664.7 E(): 0
99.678% identity (99.678% ungapped) in 1863 nt overlap (76-1938:1-1863)

```
      50      60      70      80      90     100
EP0111 CGTGTTGCGCAGCACATCTGAGGATGATTCATGAGTAAGAGACAGAGCTCGGTGCAGATC
      :
GSN:AA      ATGAGTAAGAGACAGAGCTCGGTGCAGATC
              10      20      30
```

```
     110     120     130     140     150     160
EP0111 AGGCAGCTGTTTCGGACCAGCACTCAGATCCAGGATCAGCCCAGCCTCCTCAGAGCTGGAG
      :
GSN:AA AGGCAGCTGTTTCGGACCAGCACTCAGATCCAGGATCAGCCCAGCCTCCTCAGAGCTGGAG
          40      50      60      70      80      90
```

```
     170     180     190     200     210     220
EP0111 ACCCTCTCCCCACCTCGCCTCTCGCCCCGTGACCCCCCTCGGTGACATGTACCTGAAGAG
      :
GSN:AA ACCCTCTCCCCACCTCGCCTCTCGCCCCGTGACCCCCCTCGGTGACATGTACCTGAAGAG
          100     110     120     130     140     150
```

```
     230     240     250     260     270     280
EP0111 AGCCGGGGTTTCTGGAGGGGTGGCTGCTGTGGACTTTTGGGAAGGGACGTACGACTATGCC
      :
GSN:AA AGCCGGGGTTTCTGGAGGGGTGGCTGCTGTGGACTTTTGGGAAGGGACGTACGACTATGCC
          160     170     180     190     200     210
```

```
     290     300     310     320     330     340
EP0111 GCCCCCAACCCTGCCACGACTCCCTTTACAGCCAGTCCAGCACCGGCTACTACTCTGCT
      :
GSN:AA GCCCCCAACCCTGCCACGACTCCCTTTACAGCCAGTCCAGCACCGGCTACTACTCTGCT
          220     230     240     250     260     270
```

```
     350     360     370     380     390     400
EP0111 CCCCTGGAAACAAACGGACCCCCCTCAGAAGGCAGTCTGCAGTCCCTGGGCAGTGGGCGG
      :
GSN:AA CCCCTGGAAACAAACGGACCCCCCTCAGAAGGCAGTCTGCAGTCCCTGGGCAGTGGGCGG
          280     290     300     310     320     330
```

```
     410     420     430     440     450     460
EP0111 ACGAGCCCTCTGGTGTGTTGTGCCCTCCAGCCCCAGACTCAGTCCCTTTATGCATCCACCC
      :
GSN:AA ACGAGCCCTCTGGTGTGTTGTGCCCTCCAGCCCCAGACTCAGTCCCTTTATGCATCCACCC
          340     350     360     370     380     390
```

```
     470     480     490     500     510     520
EP0111 AGCCACCACTATCTGGAAACCACTTCCACGCCCCGTTTACAGATCCAGCCACCAGGGAGCC
      :
GSN:AA AGCCACCACTATCTGGAAACCACTTCCACGCCCCGTTTACAGATCCAGCCACCAGGGAGCC
          400     410     420     430     440     450
```

```
     530     540     550     560     570     580
EP0111 TCCAGGGAGGACCACTGCGGCTCCCGGGAGGACACGTGCAGCCTGGGGGAGTTAGGCGCC
      :
GSN:AA TCCAGGGAGGACCACTGCGGCTCCCGGGAGGACACGTGCAGCCTGGGGGAGTTAGGCGCC
          460     470     480     490     500     510
```

```
     590     600     610     620     630     640
EP0111 GGAGCCGGGGCTGGGGGGTTTGAGATGGCCAAAGACACGCGTTTCTGCGCCGTGTGCAGC
      :
GSN:AA GGAGCCGGGGCTGGGGGGTTTGAGATGGCCAAAGACACGCGTTTCTGCGCCGTGTGCAGC
          520     530     540     550     560     570
```

```
     650     660     670     680     690     700
```

BNSDOCID: YF 2181483A 1

ENCLOSURE YB 2161423A 1

1970 1980 1990 2000 2010 2020
EP0111 GGCTCAAGGGTTCAGGTTGGGACAAGGTGATGCTTGATTTAATTTTAAGAATTATTATA

1 Yell
 translation of seq 1
 VS
 Seq 1A3 of SP: 20168

ID AAB20898 standard; Protein; 620 AA.
 XX
 AC AAB20898;
 XX
 DT 05-JAN-2001 (first entry)
 XX
 DE Oryzias lapites oestrogen receptor protein SEQ ID NO:3.
 XX
 KW Oryzias lapites; oestrogen receptor.
 XX
 OS Oryzias lapites.
 XX
 PN JP2000201688-A.
 XX
 PD 25-JUL-2000.
 XX
 PF 06-APR-1999; 99JP-0098787.
 XX
 PR 10-NOV-1998; 98JP-0319465.
 XX
 PA (SUMO) SUMITOMO CHEM CO LTD.
 XX
 DR WPI; 2000-567950/53.
 DR N-PSDB; AAA92175.
 XX
 PT An estrogen receptor gene and its application
 XX
 PS Claim 2; Page 13-15; 23pp; Japanese.
 XX

CC The present sequence represents an oestrogen receptor derived from
 CC Oryzias lapites. Also described are: (1) a vector comprising the
 CC oestrogen receptor gene; (2) a transformant prepared by introducing
 CC the oestrogen receptor gene or vector from (1) into a host cell;
 CC (3) a method for the preparation of an oestrogen receptor comprising
 CC culturing the transformant from (2) to produce the oestrogen receptor;
 CC and (4) a method for the evaluation of oestrogen receptor-activating
 CC ability of a chemical substance in which the chemical substance is
 CC reacted with a transformant prepared by introducing a reporter gene
 CC connected downstream of a transcription controlling region containing
 CC an oestrogen response sequence and the above oestrogen receptor gene to
 CC an oestrogen-nonendogenous host cell. The transformant can be used for
 CC the evaluation of oestrogen receptor-activating ability of a chemical
 CC substance.

XX
 SQ Sequence 620 AA;
 SQ 39 A; 44 R; 10 N; 29 D; 0 B; 18 C; 27 Q; 31 E; 0 Z; 60 G; 19 H;
 SQ 24 I; 67 L; 25 K; 20 M; 15 F; 43 P; 67 S; 30 T; 4 W; 20 Y; 28 V;
 SQ 0 Others;
 mskrqssvqi rqlfgpalrs rispassele tisprrlspr dplgdmyppe srgsggvaav
 dllegtydya apnpattply ~~sqsstgyysa pletnppse qslqslgsgp tsplvfvss~~
 prlspfmhpp shhylettst pyvrsshqga sredqgsre dtcslgelga gagaggfema
 kdtrfcavcs dyasgyhygv ~~wscecgckaff krsiqghndy mcpatnqcti dmrrrkqca~~
 crlrkcyevg mmkggvrkdr ~~irilrrdkrr tgvqgdgkvv kqgehktvhy dgrkrsstgg~~
 gggggggrls vtsippeqvl ~~lllqgaoppi lcarqqlary ytevtmtll tsmadkelvb~~
 miawakkllpg flqlslhdqv lllesswlev lmigliwrsi hcpgklifag dlildrnegd
 cvegmteifd mllatasrfr ~~vklkpeefv cikarillns gsfstctgtm eplhmsaavq~~
 smldtitdal ihyisqgyl aqeqarrqag pllllshirh msnkgmehly smkcknkvpl
 ydlillemla hrlhhpvrp ~~qslsqvdrdp pstssggggi apgsisasrg riespsrgpf~~
 apsvlqyggs rpdctpalqd

//

>>GSP:AAB20898 Oryzias latipes oestrogen recept (620 aa)
 initn: 4198 initl: 4198 opt: 4198 Z-score: 3093.7 bits: 584.7 E(): 2.3e-164
 Smith-Waterman score: 4198; 99.516% identity (99.516% ungapped) in 620 aa overlap
 (75-1934:1-620)

```

      90      120      150      180      210      240
EP0111 MSKRQSSVQIRQLFGPALRSRISPASSELETLSPPRLSPRDPLGDMYPFEESRGSGGVA AV
      .....
GSP:AA MSKRQSSVQIRQLFGPALRSRISPASSELETLSPPRLSPRDPLGDMYPFEESRGSGGVA AV
      10      20      30      40      50      60

      270      300      330      360      390      420
EP0111 DFLEGTYDYAAPNPATTPLYSQSSTGYYSAPLETNGPPSEGLQSLGSGPTSPLVFPVSS
      .....
GSP:AA DLLEGTYDYAAPNPATTPLYSQSSTGYYSAPLETNGPPSEGLQSLGSGPTSPLVFPVSS
      70      80      90      100      110      120

      450      480      510      540      570      600
EP0111 PRLSPFMHPPSHHYLETTSTPVYRSSHQGASREDQCGSREDTCSLGELGAGAGAGGFEMA
      .....
GSP:AA PRLSPFMHPPSHHYLETTSTPVYRSSHQGASREDQCGSREDTCSLGELGAGAGAGGFEMA
      130      140      150      160      170      180

      630      660      690      720      750      780
EP0111 KDTRFCAVCSDYASGYHYGVWSCEGCKAFFKRSIQGHNDYMCPATNQCTIDNRNRKSCQA
      .....
GSP:AA KDTRFCAVCSDYASGYHYGVWSCEGCKAFFKRSIQGHNDYMCPATNQCTIDNRNRKSCQA
      190      200      210      220      230      240

      810      840      870      900      930      960
EP0111 CRLRKCYEVGMMKGGVRKDRIRILRRDKRRTGVGDGDKVVKQEHKTVHYDGRKRSSTGG
      .....
GSP:AA CRLRKCYEVGMMKGGVRKDRIRILRRDKRRTGVGDGDKVVKQEHKTVHYDGRKRSSTGG
      250      260      270      280      290      300

      990      1020      1050      1080      1110      1140
EP0111 GGGGGGGRLSVTSIPPEQVLLLLQGAEPFILCSRQKLSRPYTEVTMMTLLTSMADKELVH
      .....
GSP:AA GGGGGGGRLSVTSIPPEQVLLLLQGAEPFILCSRQKLSRPYTEVTMMTLLTSMADKELVH
      310      320      330      340      350      360

      1170      1200      1230      1260      1290      1320
EP0111 MIAWAKKLPGFLQSLHDQVLLLESSWLEVLMIGLIWRSIHCPGKLIFAQDLILDRNEGD
      .....
GSP:AA MIAWAKKLPGFLQSLHDQVLLLESSWLEVLMIGLIWRSIHCPGKLIFAQDLILDRNEGD
      370      380      390      400      410      420

      1350      1380      1410      1440      1470      1500
EP0111 CVEGMTEIFDMLLATASRFRVLKLPKEEFVCLKAIILLNSGAFSFCGTGTMEPLHNSAAVQ
      .....
GSP:AA CVEGMTEIFDMLLATASRFRVLKLPKEEFVCLKAIILLNSGAFSPCTGTGTMEPLHNSAAVQ
      430      440      450      460      470      480

      1530      1560      1590      1620      1650      1680
EP0111 SMLDTITDALIHYSISQGYLAQEQAARQAQLLLLLSHIRHMSNKGMEHLYSMKCKNKVPL
      .....
GSP:AA SMLDTITDALIHYSISQGYLAQEQAARQAQLLLLLSHIRHMSNKGMEHLYSMKCKNKVPL
      490      500      510      520      530      540

      1710      1740      1770      1800      1830      1860
EP0111 YDLLLEMLDAHRLHHPVRAPQSLSQVDRDPPSTSSGGGGIAPGSISASRGRIESPSRGPF
      .....
GSP:AA YDLLLEMLDAHRLHHPVRAPQSLSQVDRDPPSTSSGGGGIAPGSISASRGRIESPSRGPF
      550      560      570      580      590      600

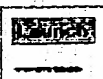
```

01000010 10 01010011 1

This entry is from:
SWALL (SPTR)

Save

Link



Print Entry Summary

General Description References Comments Links Keywords Features Sequence

General information

Entry name ESR1_ORYLA
Accession number P50241
Created Rel. 34, 1-OCT-1996
Sequence update Rel. 37, 15-DEC-1998
Annotation update Rel. 40, 16-OCT-2001

01-10-1996

1-5

5

Description and origin of the Protein

Description ESTROGEN RECEPTOR (ER) (ESTRADIOL RECEPTOR) (ER-ALPHA).
Gene name(s) ESR OR NR3A1 OR MER.
Organism source Oryzias latipes (Medaka fish).
Taxonomy Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi; Actino-
Neopterygii; Teleostei; Euteleostei; Neoteleostei; Acanthomorpha; Acantho-
Percomorpha; Atherinomorpha; Beloniformes; Adrianichthyidae; Oryziinae;
NCBI TaxID 8090

References

- [1] Okada, H., Kawahara, T., Yamashita, I., RL Submitted (APR-1998) to the EMBL/GenBank/DDBJ databases.
Position RP SEQUENCE FROM N.A.
Comments RC STRAIN=D-RR; TISSUE=LIVER;
- [2] Kawahara, T., Yamashita, I., RT "Oryzias latipes genomic DNA for estrogen receptor. RL Submitted to the EMBL/GenBank/DDBJ databases.
Position RP SEQUENCE FROM N.A.

Comments

FUNCTION THE STEROID HORMONES AND THEIR RECEPTORS ARE INVOLVED IN THE REGULATION OF EUKARYOTIC GENE EXPRESSION AND A CELLULAR PROLIFERATION AND DIFFER IN TARGET TISSUES.

SUBUNIT BINDS DNA AS A HOMODIMER. CAN FORM HETERODIMER WITH ER- BETA (BY SIMILARITY).

SUBCELLULAR LOCATION NUCLEAR.

DOMAIN COMPOSED OF THREE DOMAINS: A MODULATORY N-TERMINAL DOMAIN, A DNA-BINDING DOMAIN, AND A C-TERMINAL STEROID-BINDING DOMAIN.

SIMILARITY BELONGS TO THE NUCLEAR HORMONE RECEPTOR FAMILY. NR3 SUBFAMILY.

Copyright

This SWISS-PROT entry is copyright. It is produced through a collaboration between the Swiss Bioinformatics and the EMBL outstation - the European Bioinformatics Institute. There are no restrictions on its use by non-profit institutions as long as its content is in no way modified and this statement

removed. Usage by and for commercial entities requires a license agreement (See <http://www.isb-sib.ch/announce/> or send an email to license@isb-sib.ch).

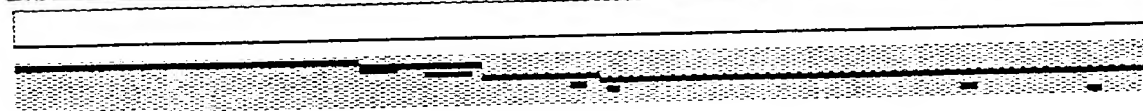
Database cross-references

EMBL	D28954;BAA25900.1;-.
	AB033491;BAA86925.1;-.
HSSP	P03372;1HCP.
	IPR000536;Hormone_rec_lig.
	IPR001292;Oest_recep.
InterPro	IPR001723;Strdhormone_receptor.
	IPR001628;zf-C4.
	PF00104;hormone_rec;1.
Pfam	PF02159;Oest_recep;1.
	PF00105;zf-C4;1.
PRINTS	PR00398;STRDHORMONER.
	PR00047;STROIDFINGER.
SMART	SM00430;HOLI;1.
	SM00399;ZnF_C4;1.
PROSITE	PS00031;NUCLEAR_RECEPTOR;1.

Keywords

Receptor, Transcription regulation; DNA-binding; Nuclear protein; Zinc-finger; Steroid-binding;

Features



Key	Begin	End	Length	Description
DOMAIN	1	185	185	MODULATING.
DNA_BIND	186	251	66	NUCLEAR RECEPTOR-TYPE.
ZN_FING	186	206	21	C4-TYPE.
ZN_FING	222	246	25	C4-TYPE.
DOMAIN	252	314	63	HINGE.
DOMAIN	315	620	306	STEROID-BINDING.
DOMAIN	299	307	9	POLY-GLY.
DOMAIN	320	323	4	POLY-LEU.
DOMAIN	511	515	5	POLY-LEU.
DOMAIN	576	579	4	POLY-GLY.

Sequence information

Length: 620 aa, molecular weight: 67729 Da, CRC64 checksum: DDCBD18C2B2BA522

MSKRQSSVQI RQLFGPALRS RISPASSELE TLSPPRLSPR DPLGDMYPEE SRGSGGVA
DFLEGTYDYA APNPATTPLY SQSSTGYISA PLETNGPPSE GSLQSLGSGP TSPLVFVP
PRLSPFMHPP SHHYLETTST PVYRSSHQGA SREDQCGSRE DTCSLGELGA GAGAGGF
KDTRFCAVCS DYASGYHYGV WSCGCKAFF KRSIQGHNDY MCPATNQCTI DRNRKRS
CRLRKCYEVG MMKGGVRKDR IRILRRDKRR TGVGDGDKV KGQEHKTVHY DGRKRS
GGGGGGGRLS VTSIPPEQVL LLLQGAEPPI LCSRQKLSRP YTEVTMMTLL TSMADKELV
MIAWAKKLPGL FLQLSLHDQV LLESSWLEV LMIGLIWRSI HCPGKLIFAQ DLILDRNEGD

CVEGMTEIFD MLLATASRFR VLKLPKEEFV CLKAIILLNS GAFSFCTGTM EPLHNSAAVQ
SMLDTITDAL IHYISQSGYL AQEQARRQAQ LLLLLSHIRH MSNKGMEHLY SMKCKNKVPI
YDLLLEMLDA HRLHHPVRAP QSLSQVDRDP PSTSSGGGGI APGSISASRG RIESPSRGF
APSVLQYGGG RPDCTPALQD 620

//

General	Description	References	Comments	Links	Keywords	Features	Sequence
---------	-------------	------------	----------	-------	----------	----------	----------

SRS 6.1.3 | feedback

BNSDOCID <XP 21B1486A 1 >

発送番号 1 2 1 2 6 4

発送日 平成 1 5 年 4 月 1 5 日 3 / 3

・ 先行技術文献

Winn R., Marine Environmental Research, vol. 46 (1-5), p. 130 (1998)

Takagi S. et al., Molecular Marine Biology and Biotechnology, vol. 3 (4),

pp. 192-199 (1994)

Gray M. A. et al., Environmental Toxicology and Chemistry, vol. 18 (11),

pp. 2587-2594 (1999)

この先行技術文献調査結果の記録は、拒絶理由を構成するものではない。

Aryl Hydrocarbon Receptor is Required for Prevention of Blood Clotting and for the Development of Vasculature and Bone in the Embryos of Medaka Fish, *Oryzias latipes*

Toshiyuki Kawamura and Ichiro Yamashita*

Center for Gene Science, Hiroshima University, Kagamiyama 1-4-2,
Higashi-Hiroshima 739-8527, Japan

ABSTRACT—The aryl hydrocarbon receptor (AHR) is a member of ligand-activated transcription factors and conserved among vertebrates. To investigate the role of AHR in fish development, medaka embryos were treated with agonist (2,3,7,8-tetrachlorodibenzo-*p*-dioxin), antagonists (α -naphthoflavone and resveratrol), and inhibitor (piperonyl butoxide) of cytochromes (Cyts) P450 encoded by a battery of target genes. These embryos were found to have similar abnormal phenotypes. Among the most consistent phenotypes were blood clotting and malformation of bone that were associated with vascular damages. These results thus indicate that control of AHR is important for proper development of fish embryos. AHR may control levels of Cyts P450 that are responsible for synthesis and metabolism of a toxic compound that caused the abnormal phenotypes. Complementary DNA fragments encoding AHR homologs were cloned from medaka embryos. AHR-specific mRNA was ubiquitously expressed in embryos and adult tissues.

Key words: aryl hydrocarbon receptor, blood clotting, bone formation, cytochrome P450, dioxin.

INTRODUCTION

Planar halogenated hydrocarbons, such as 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD), are notorious environmental pollutants that are extremely toxic to early stages of vertebrate development (Peterson *et al.*, 1993). Hallmark signs of TCDD toxicity in fish sac fry are yolk sac edema, slowed blood flow, hemorrhage, and growth retardation culminating in mortality (Cantrell *et al.*, 1996; Henry *et al.*, 1997; Hornung *et al.*, 1999). Vascular damage, as assessed by TCDD-induced apoptotic cell death, is a key physiological mediator of the embryo toxicity (Cantrell *et al.*, 1996; Cantrell *et al.*, 1998). These chemicals bind to a ligand-dependent transcriptional factor called the aryl hydrocarbon receptor (AHR), resulting in the activation of a battery of genes encoding various cytochromes (Cyts) P450 that are responsible for degradation of the environmental contaminants (Hankinson, 1995; Guiney *et al.*, 1997; Guiney *et al.*, 2000). AHR is conserved among vertebrates, thus, may have arisen in an ancestral vertebrate as a detoxification system.

Although, to date, an endogenous ligand for AHR has not been found, AHR is ubiquitously expressed in most

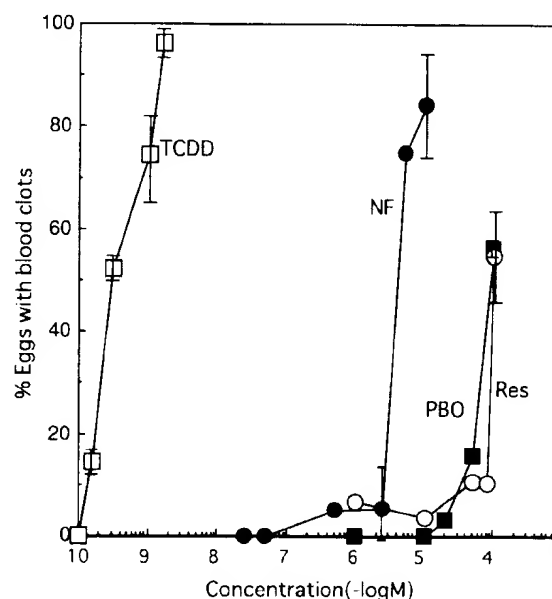
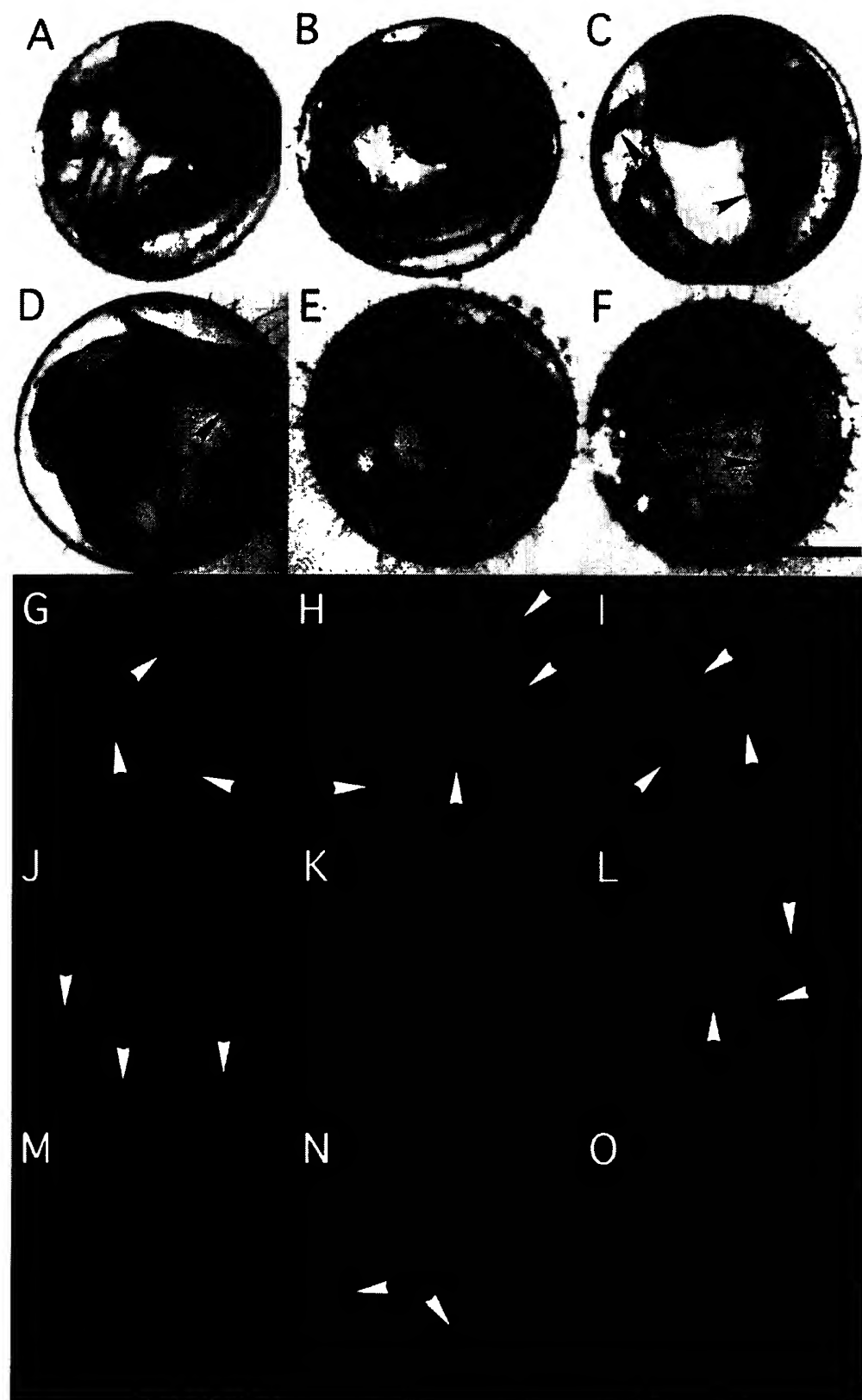


Fig. 1. Effects of 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD), α -naphthoflavone (NF), resveratrol (Res), and piperonyl butoxide (PBO) on blood clotting during the embryo stage. Eggs were treated with TCDD, NF, Res, or PBO at the indicated concentrations until 6, 6, 4, or 5 dpf, respectively, and counted for blood clots.

* Corresponding author: Tel. +81-824-24-6271;
FAX. +81-824-22-7184.
E-mail: iyama@hiroshima-u.ac.jp



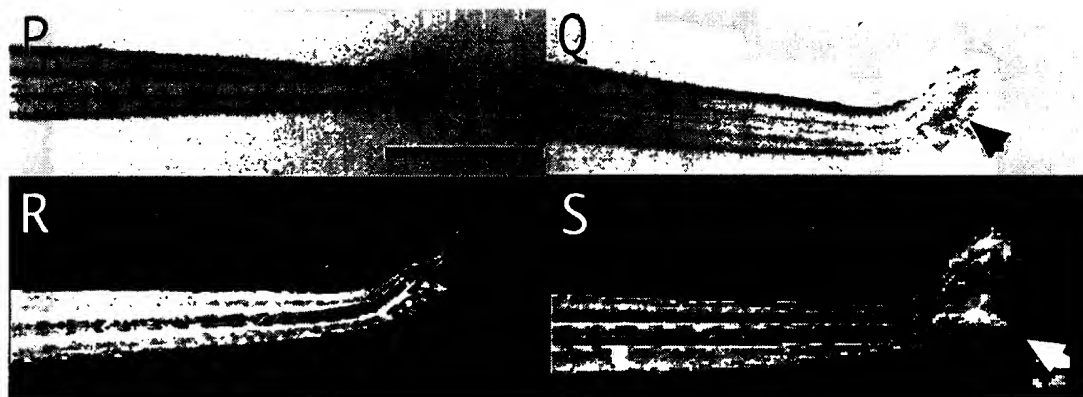


Fig. 2. Photographs of blood clots, yolk vein, and fin. Eggs and fry were treated with 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD), α -naphthoflavone (NF), resveratrol (Res), or piperonyl butoxide (PBO) as follows and photographed for blood clots (A–F), yolk vein under green fluorescence (G–O), and fin (P–S): (A) mock-treated, 5 dpf; (B, C) 1.55 nM TCDD, 5 and 7 dpf; (D) 10 μ M NF, 5 dpf; (E) 100 μ M PBO, 5 dpf; (F) 100 μ M Res, 4 dpf; (G–I) mock-treated at 3, 5, and 7 dpf; (J, K) 1.55 nM TCDD, 5 and 7 dpf; (L, M) 10 μ M NF, 3 and 5 dpf; (N, O) 100 μ M PBO, 4 and 5 dpf; (P, R) mock-treated, 5-day post-hatching; and (Q, S) 0.155 nM TCDD, 5-day post-hatching. Arrows indicate blood clots (B–F, and Q), yolk veins (G–J, L, and N), and the constricted fin (S). Bar, 0.5 mm.

organs and cells in the body (Rowlands and Gustafsson, 1997). However, there is only a limited knowledge of developmental and physiological functions of AHR in the mouse (Gonzalez and Fernandez-Salguero, 1998), although the role of AHR in detoxification of environmental aryl hydrocarbons has been extensively studied *in vitro* (Hankinson, 1995). AHR-null mice were resistant to the acute toxicity (Fernandez-Salguero *et al.*, 1996) of and the teratogenic response (Mimura *et al.*, 1997) to TCDD, and found to have a number of abnormal phenotypes such as decreased accumulation of lymphocytes in the spleen and lymph nodes and reduction in liver size that are associated with accelerated rates of apoptosis (Fernandez-Salguero *et al.*, 1995), and difficulties in reproduction (Abbott *et al.*, 1999; Robles *et al.*, 2000). Thus, AHR is involved in the toxicity of and the teratogenesis by TCDD *in vivo*, and plays an important role in the development of the liver and the immune system, and in reproduction. However, no such function has been elucidated in other vertebrates.

Here we re-evaluated the role of AHR in chemical toxicity of TCDD in medaka fish embryos because there have been no pharmacological studies in fish using antagonist and also examined for any possible developmental and physiological function of AHR in medaka fish embryos using antagonists and Cyts P450 inhibitor. We found that AHR mediates TCDD toxicity such as blood clotting, malformation of bone, and regression of blood vessels, and that AHR is required for the embryonic development of vasculature and bone. To our knowledge, this is the first report of the developmental role of AHR in lower vertebrates.

MATERIALS AND METHODS

Fish and embryo culture

We used the d-rR strain of medaka fish, *O. latipes* (Kawahara and Yamashita, 2000). The fish were maintained at 25–26°C under

artificial photo-period of 14L 10D, and fed by powdered TetraMin (Tetra). Eggs were collected within 12 hr postfertilization (hpf), rinsed with tap water, and immersed in Yamamoto's salt solution (Yamamoto, 1969) with or without test chemicals. At least 30 eggs were used in each experiment. TCDD was purchased from Cambridge Isotope Laboratories, Inc. Antagonists, α -naphthoflavone (NF) (Gasiewicz and Rucci, 1991; Merchant *et al.*, 1993) and resveratrol (Res) (Ciolino *et al.*, 1998; Casper *et al.*, 1999; Singh *et al.*, 2000), were from Sigma. Cyts P450 inhibitor, piperonyl butoxide (PBO) (Dahl and Hodgson, 1979; Testa and Jenner, 1981; Adams *et al.*, 1993), was from Tokyo Kasei Kogyo Co. These reagents were dissolved in acetone. The stock solutions were diluted over 1,000-fold with Yamamoto's solution and added to eggs of 12 hpf for NF, Res, and PBO or of 24 hpf for TCDD. The solvent was added to the mock-treated eggs as a control. The reducing agent, N-acetyl cysteine (NAC) (Sigma), was dissolved in Yamamoto's solution and added to 12 hpf eggs. Eggs and fry were cultured under the same condition as above (except without feed) and inspected for blood clotting under a dissecting microscope. Eggs and fry in which blood clots formed were counted.

Data are presented as mean \pm SEM. Statistical significance between values of control and experiment was assessed by Student's *t*-test.

Observation of blood vessels

In order to observe the development of blood vessels, eggs were fixed with 4% paraformaldehyde for 3 days and observed under green fluorescence with a filter set (excitation filter, 546/10 nm, barrier filter, 590 nm) in Leica MZ FLIII stereo-fluorescence microscope. The fixed eggs were also dechorionated with forceps and stained with hematoxylin.

Bone staining

In order to observe the bone development, calcified bone was stained with alizarin S essentially as described (Takeuchi, 1960). In brief, fish were anesthetized with 0.015% phenylurethane, skinned with forceps, treated with 2% KOH for 24 hr, and finally stained with 0.1% alizarin S solution. After washing in tap water, the fish were successively transferred to 50%, and 70%, and finally embedded in 100% glycerin. Anesthetized fry were directly treated with 2% KOH for 2 h, fixed in 4% paraformaldehyde for 24 hr, then stained with alizarin S.

Isolation of cDNAs encoding medaka AHR homologs

As PAS domain of AHR is highly conserved among vertebrates (Rowlands and Gustafsson, 1997), a corresponding region of cDNA was amplified with degenerated oligonucleotides (AhR-A1 and AhR-B1) as described (Hahn and Karchner, 1995) using total RNA from 6-day postfertilization (dpf) medaka embryos. The cDNA fragment was cloned in plasmid and sequenced. Based on the sequence, nested oligonucleotides were designed and 5' and 3' RACEs (rapid amplification of cDNA ends) were performed on the same RNA by using 5' and 3' RACE Systems (GIBCO BRL), yielding the remainder of the coding sequence, 5' and 3' untranslated regions, and polyadenylation sequence.

RNA analysis

Total RNA was extracted from embryos and adult tissues as described (Kawahara *et al.*, 2000). RT-PCR (reverse transcription-polymerase chain reaction) analysis was done as described (Kawahara *et al.*, 2000) with the primers as follows for generation of the 437-bp cDNA encoding a part of PAS domain: poly(dT) oligonucleotide used for RT, and 5'-CCAGCAGGAGTTCAGGAGGA and 5'-ATTTACCCTTTGCGTCACA for PCR. Amplified DNA was electrophoresed in 1% agarose gel and stained with ethidium bromide.

RESULTS

AHR mediates the toxic effects of TCDD on vascular development

We re-evaluated the toxic effects of TCDD on medaka embryos. To do this, embryos (1 dpf) were immersed in saline solution for medaka containing increasing concentrations of TCDD, and observed for any abnormal phenotype under a dissecting microscope (Fig. 1). Clearly visible signs of blood clotting were apparent after 4 days in caudal veins of TCDD-treated embryos (Fig. 2B), although blood cells were circulating in vasculature (Fig. 2J) but at a reduced rate. Blood clots were also found in yolk veins after 6 days (Fig. 2C), at that time, vascular structure was almost absent (Fig. 2K). In control embryos, yolk veins were apparent at 3 dpf (Fig. 2G) and developed progressively in a curve structure (Fig. 2A, H and I). Very small blood clots were occasionally found in yolk veins of normal embryos (less than 3%), but not scored in this study. These results are consistent with the previous observation that TCDD induces apoptosis of blood vessels (Cantrell *et al.*, 1996).

If TCDD induced the vascular damage through activation of AHR, the antagonist (NF) would reduce the extent to which blood clotting was detected. For this purpose, two different experiments were done, in which embryos were treated with high (1.55 nM) or medial (0.775 nM) concentration of TCDD (Fig. 3A or B, respectively). For both cases, addition of NF effectively suppressed blood clotting but only transiently (Fig. 3A and B). However, in the latter case, NF markedly enhanced the hatching success of TCDD-treated embryos, giving rise to almost complete hatching (Fig. 3C). These results indicate that TCDD-induced vascular damage is mediated through activation of AHR.

It is well known that TCDD-bound AHR activates transcription of a battery of genes encoding Cyts P450. If these

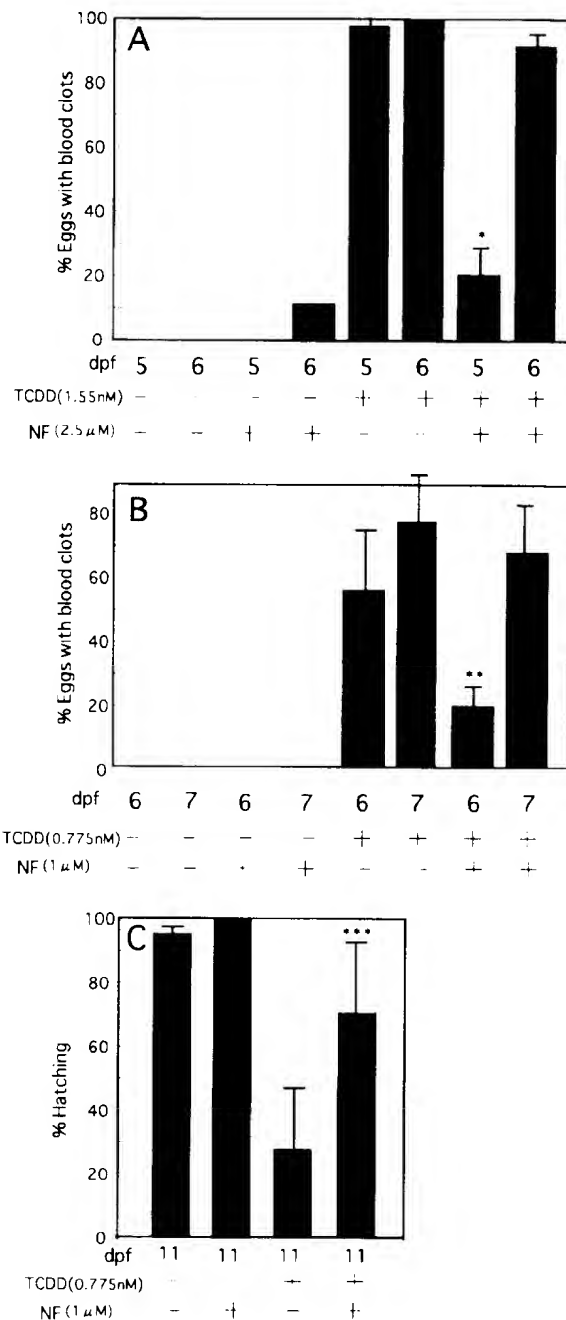


Fig. 3. Suppression by α -naphthoflavone (NF) of 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD)-induced blood clotting and mortality. (A) Eggs were treated with 1.55 nM TCDD and 2.5 μ M NF until 5 and 6 dpf as indicated, and examined for blood clotting. * $P < 0.01$. (B) Eggs were treated with 0.775 nM TCDD and 1 μ M NF until 6 and 7 dpf as indicated, and examined for blood clotting. ** $P < 0.2$. (C) Eggs were treated as described in (B), and examined for hatching rate at 11 dpf. *** $P < 0.05$.

enzymes were involved in the TCDD-induced toxicity, an inhibitor of P450 would reduce the rate of TCDD-induced blood clotting. We therefore examined the ability of PBO to provide protection against high concentration (1.55 nM) of

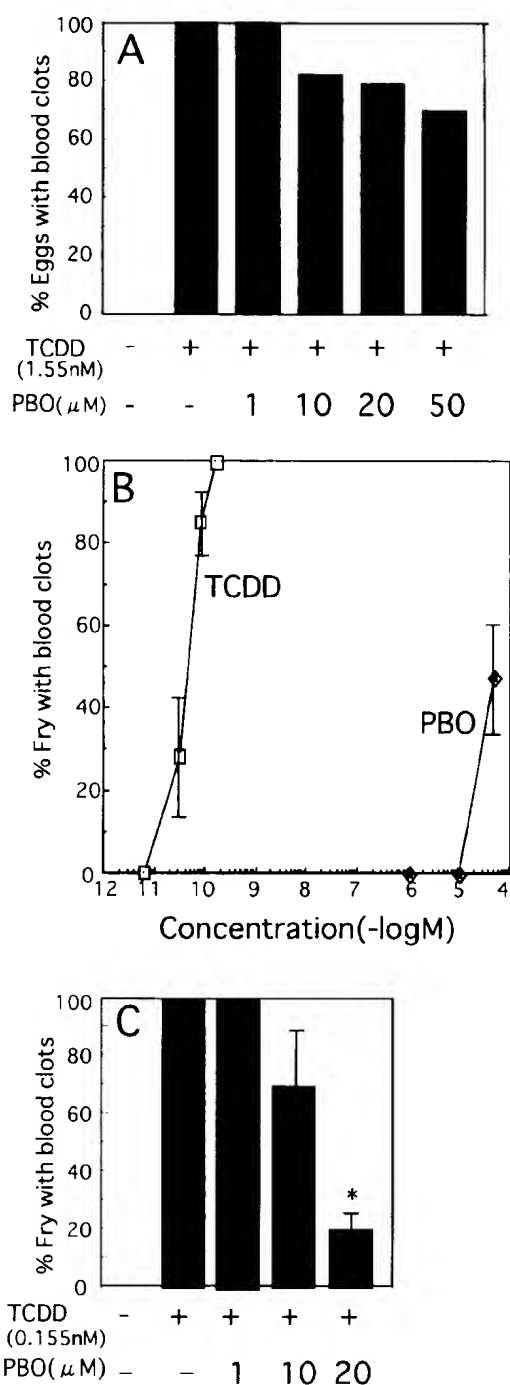


Fig. 4. Suppression by piperonyl butoxide (PBO) of 2,3,7,8-tetra-chlorodibenzo-*p*-dioxin (TCDD)-induced blood clotting. (A) Eggs were treated with 1.55 nM TCDD and increasing concentrations (μM) of PBO as indicated until 5 dpf, and examined for blood clotting. (B) Eggs were treated with TCDD and PBO at the indicated concentrations until 5-day post-hatching, and examined for blood clotting in the caudal fin. (C) Eggs were treated with 0.155 nM TCDD and increasing concentrations of PBO as indicated until 5-day post-hatching, and examined for blood clotting in the caudal fin. * $P < 0.05$.

TCDD (Fig. 4A). Unexpectedly, PBO reduced the blood clotting rate only slightly; we cannot use higher concentrations of PBO because PBO itself induced blood clotting (described below). We therefore tried to seek for conditions under which lower concentrations of TCDD induce blood clotting effectively. We found that blood clots formed in the caudal fin (Fig. 2Q) after immersing embryos until 5-day post-hatching at subnanomolar concentrations of TCDD (Fig. 4B). Blood clots did not form in the control fin (Fig. 2P). Under the above condition, PBO effectively suppressed the adverse effect of TCDD (Fig. 4C). These results suggest that the TCDD-induced toxicity was caused by elevated expression of a certain Cyt P450.

Previous reports conclude that oxidative stress caused by TCDD-induced expression of Cyts P450 contributes to embryotoxicity and vascular damage associated with apoptosis, because the reducing agent, NAC, partially recovers

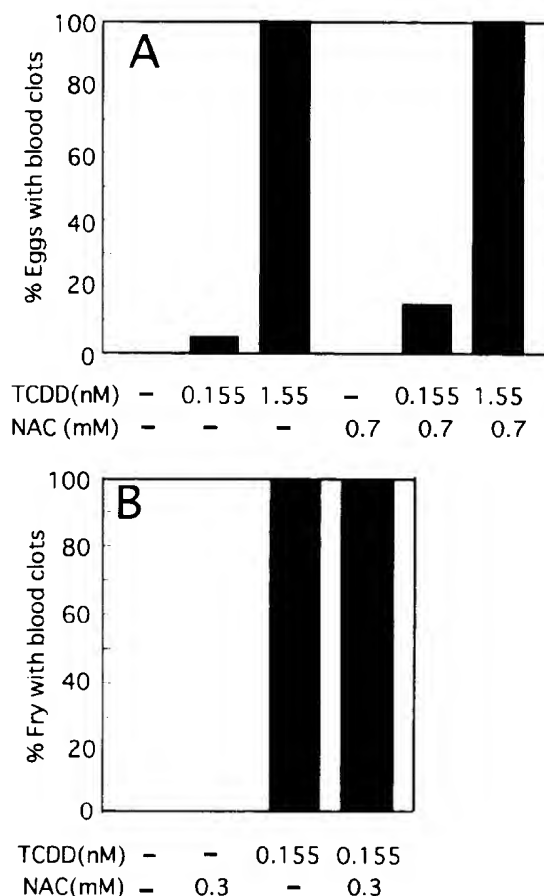


Fig. 5. N-acetyl cysteine (NAC) fails to suppress the 2,3,7,8-tetra-chlorodibenzo-*p*-dioxin (TCDD)-induced blood clotting. (A) Eggs were treated with TCDD (nM) and NAC (mM) at the indicated concentrations until 5 dpf, and examined for blood clotting. (B) Eggs were treated with 0.155 nM TCDD and 0.3 mM NAC as indicated until 5-day post-hatching, and examined for blood clotting in the caudal fin.

the TCDD-induced embryotoxicity (Cantrell *et al.*, 1996): they observed 41% survival of the embryos that had been treated with 28 nM TCDD for 2 hr and released in 0.1 mM NAC until 3 days posthatch, in contrast to 2% survival of the embryos that had been treated with TCDD and released in water. The ability of NAC to inhibit TCDD-induced toxicity was re-assessed by adding 0.7 mM (Fig. 5A) or 0.3 mM (Fig. 5B) NAC to eggs before and during the treatment with TCDD. NAC could not inhibit the blood clotting induced by 0.155 or 1.55 nM TCDD. NAC itself induced blood clotting at more than 0.9 mM (data not shown). These results suggest that general oxidative stress is not responsible for the TCDD-induced blood clotting.

Vascular damage induced by antagonists (NF and Res) and Cyts P450 inhibitor (PBO)

At the initial experiments determining the concentrations of reagents used, we found that NF, Res, and PBO induced blood clotting at higher concentrations than those used for suppression of TCDD-induced toxicity (Fig. 1). Blood clots formed in caudal and yolk veins (Fig. 2D-F). Yolk veins developed normally at the early time of incubation (up to 4 dpf) (Fig. 2L and N), but their regression was apparent at the time when blood clots formed in yolk veins (at 5 dpf) (Fig. 2M and O). These results suggest that either inactivation of AHR by NF and Res or inhibition of certain Cyts P450 by PBO caused vascular damage and blood clotting.

If the hypothesis were true, antagonist of AHR and Cyts P450 inhibitor would act synergistically to cause toxicity. We examined the synergy between low concentrations of NF (2.5 μ M) and PBO (20 μ M) that alone did not show any effect. Combination of these chemicals clearly increased the

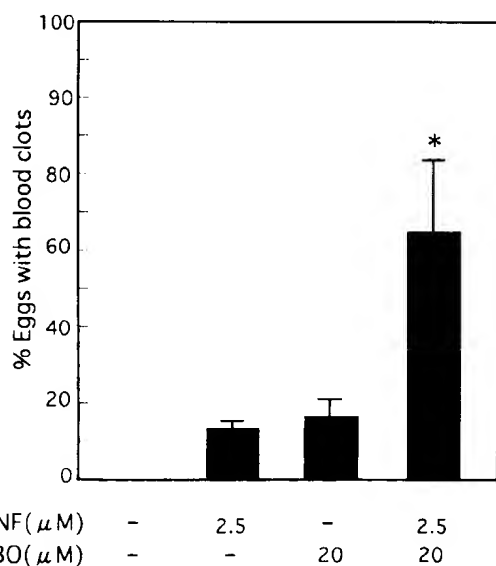


Fig. 6. Synergistic effects of α -naphthoflavone (NF) and piperonyl butoxide (PBO) on blood clotting. Eggs were treated with NF and PBO at the indicated concentrations (μ M) until 6 dpf, and examined for blood clotting. * $P < 0.2$.

rate of blood clotting (Fig. 6). We therefore conclude that control of AHR activity and levels of Cyts P450 is required for proper development of vasculature in fish.

Malformation or degeneration of bone induced by TCDD, NF, and PBO

During the experiments by incubating eggs with lower concentrations of TCDD (less than 80 pM) until 7 dpf, most eggs developed normally in appearance and blood clots did not form. The eggs were transferred to Yamamoto's solution, then to aquaria after hatching, and reared to adult by normal diet as usual. Unexpectedly we found that these fish were deformed in shape like wavy mutants (Takeuchi, 1960). We examined the bone development by staining with alizarin S. The vertebral column of TCDD-treated fish curved dorso-ventrally and laterally (Fig. 7A and B). Neural and haemal spines were short in length and deformed (Fig. 7B). NF also suppressed the TCDD-induced toxicity on bone formation (Fig. 7C), indicating the involvement of AHR.

We examined the effect of TCDD on the embryonic bone formation by incubating eggs with TCDD until 5 days post-hatching. The staining of the fry with alizarin revealed the absence of calcification in the posterior region of spinal cord and in spines (Fig. 7D and E). We also found that caudal fins were round in shape and constricted (indicated by arrow in Fig. 2S) in the TCDD-treated fry.

In order to examine the possible function of AHR and Cyts P450 in the embryonic bone formation, eggs were treated with NF or PBO until 5 days post-hatching. The treatment with NF (2.5 μ M) did not cause blood clotting in any portion of the fry (data not shown), which was different from the result with TCDD (Figs. 2Q and 4B). However, the treatment also caused degeneration of the posterior end of the spinal cord, but with normal development of spines (Fig. 7D, data not shown). PBO (50 μ M) also caused the same defect in bone formation as that NF did (data not shown).

We further examined whether NF affects homeostasis of adult fish. To do this, adult fish which had been reared by normal diet for 2 months were fed by NF-containing diet (2 mg NF/g diet) for 2 months. During the cultivation, population of fish lacking posterior fins including anal, caudal, and dorsal fins appeared after a month and became increasing near to 100% by two months (Fig. 7F).

Taken together, these results suggest that hyperactivation of AHR by TCDD is toxic to the embryonic development of bone and caudal fin, that AHR is required for proper development of bone and homeostasis of posterior fins, and that a certain Cyt P450 is also required for bone development.

Isolation and characterization of cDNAs encoding AHR homologs of medaka fish, and ubiquitous expression of AHR mRNA

We first obtained four independent cDNA clones (clones 1, 2, 3 and 4) corresponding to PAS domain (Fig. 8A). These clones were found to be identical by sequencing.

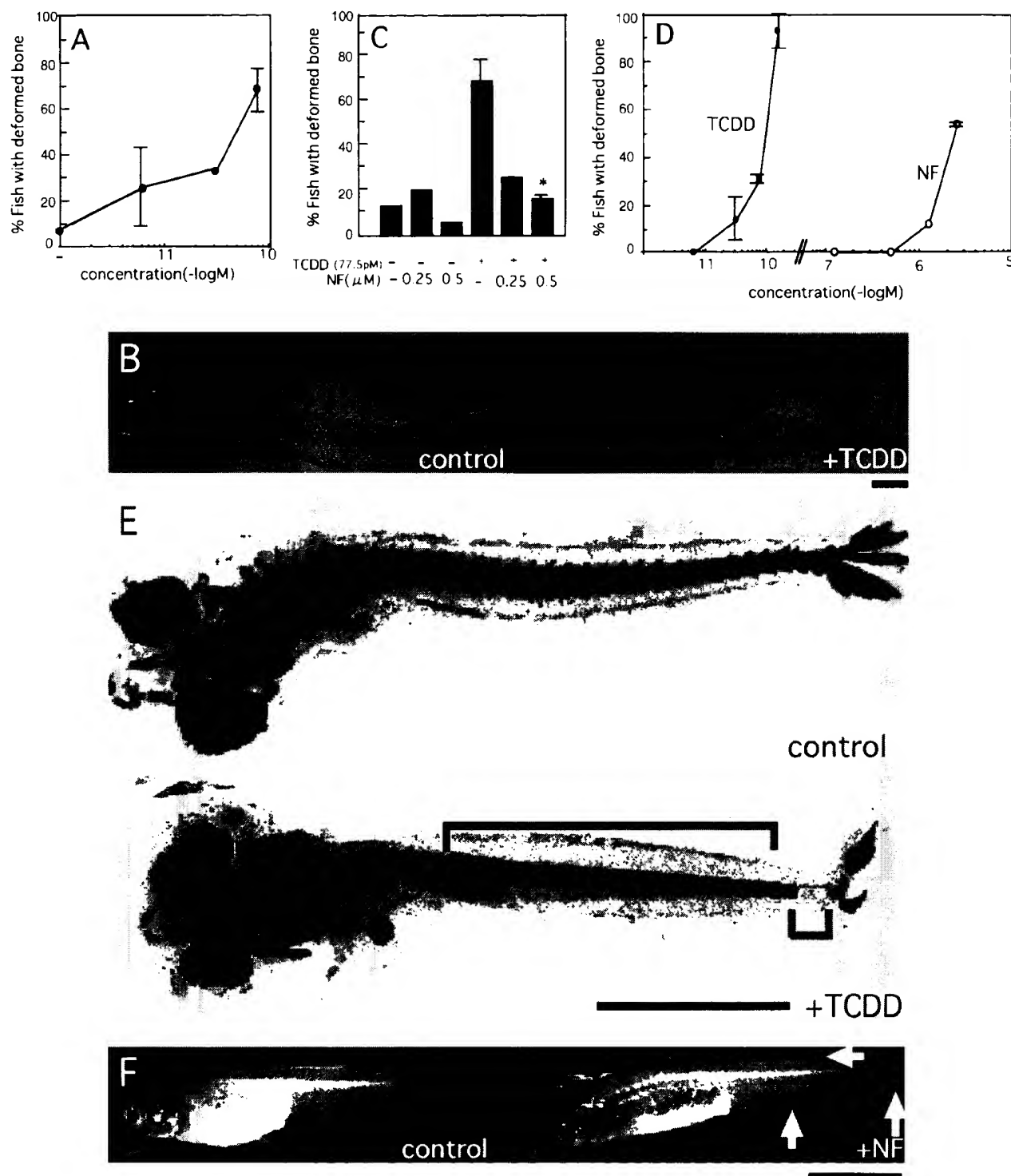


Fig. 7. Effects of 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD) and α -naphthoflavone (NF) on bone formation. (A) Eggs were treated with TCDD at the indicated concentrations until 7 dpf, and reared to adult under TCDD-free condition. The adult fish were examined for bone formation after staining with alizarin. (B) Alizarin-stained bone of mock-treated (control) and TCDD (77.5 pM)-treated fish in (A). (C) Eggs were treated with 77.5 pM TCDD and NF at the indicated concentrations (μ M) until 7 dpf, reared to adult under normal condition, and examined for bone formation. * $P < 0.05$. (D) Eggs were treated with increasing concentrations of TCDD and NF until 5-day post-hatching, and examined for bone formation. (E) Alizarin-stained bone of mock- (control) and TCDD (0.155 nM)-treated fish in (D). Spines and posterior spinal bone are absent in the TCDD-treated fry as noted. (F) Normal adult fish were fed by NF-containing diet (2 mg NF/g diet) for 2 months, and photographed. Arrows indicate the degenerated fins. Bar, 1 mm in (B) and (E), and 5 mm in (F).

Next, 5' and 3' RACEs were performed, yielding four (clones 15, 24, 27 and 30) and six (clones 307–309, 314, 315, and 319) independent clones, respectively (Fig. 8A). Four clones from 5' RACE were identical. Six clones from 3' RACE were subdivided into three identical pairs, which differ from each other only in the 3' proximal sequences denoted by broken and dotted lines in Fig. 8A. Thus, we obtained three different cDNAs, named *ahr-1*, -2, and -3 (DDBJ accession numbers AB065092, AB065093, and AB065094, respectively). However, *ahr-1* and *ahr-3* encoded the same protein (AHR1 α), and *ahr-2* encoded another homolog (AHR1 β). AHR1 α and

AHR1 β differ from each other in the C-terminal peptides (amino acid 780–879 and 780–784) denoted by shaded and dotted boxes (Fig. 8A).

AHR1 α and AHR1 β are composed of 879 and 784 amino acids with calculated molecular weights of 95.5 and 85.3 kDa, respectively. Both proteins may be classified into a type of AHR1 because they are most homologous to AHR1 of the teleost *Fundulus heteroclitus* (Karchner *et al.*, 1999) (Fig. 8B). The medaka AHR1 α and AHR1 β are also composed of three conserved domains such as basic-helix-loop-helix (bHLH), Per-ARNT-Sim (PAS), and glutamine-rich

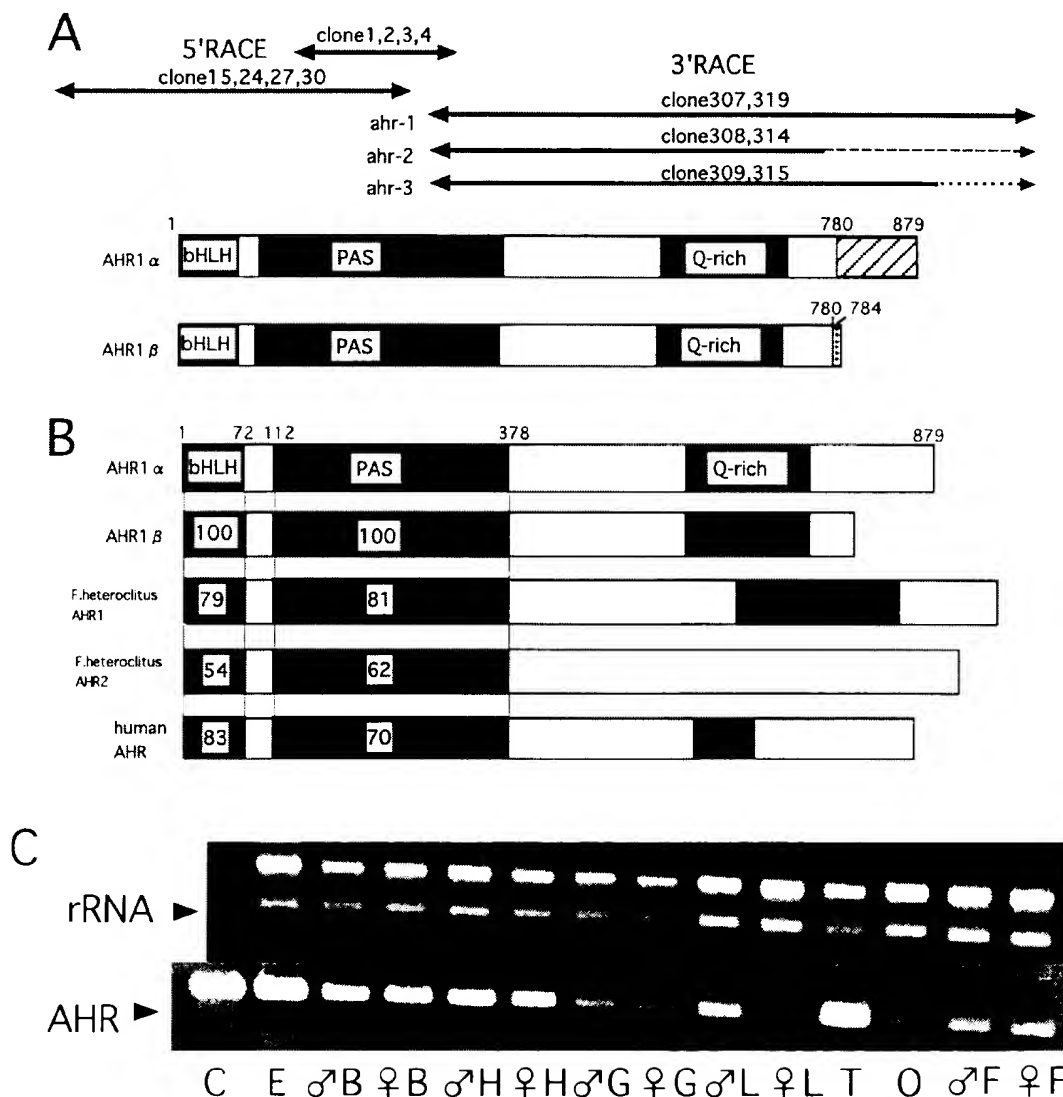


Fig. 8. Schematic drawing of the cDNAs cloned and the deduced proteins, and ubiquitous expression of AHR mRNA. **(A)** Inserts in the plasmid clones are shown on the deduced proteins (AHR1 α and AHR1 β). Plasmid numbers are marked on the corresponding inserts. The three cDNAs which differ from each other only in the 3' terminal sequences (denoted by broken and dotted lines) are named *ahr-1*, -2, and -3. AHR1 α and AHR1 β , in which three conserved domains are marked by bHLH, PAS, and Q-rich, differ from each other only in the C-terminal short peptides marked by shaded and dotted boxes. **(B)** Identity (%) of amino acid sequence among bHLH and PAS domains of AHRs from medaka, *F. heteroclitus* (killifish), and human (Dolwick *et al.*, 1993). **(C)** RT-PCR analysis of total RNAs from medaka embryos (6 dpf) and adult tissues. Symbols: B, brain; C, the control band amplified from the cDNA; E, embryo; F, caudal fin; G, gill; H, heart; L, liver; O, ovary; and T, testis. Ribosomal RNAs in the RNA samples are also shown.

(Q) domains (Rowlands and Gustafsson, 1997) (Fig. 8B).

Expression of AHR mRNA was analyzed by RT-PCR on total RNAs prepared from medaka embryos and adult tissues such as brain, fin, gill, heart, liver, ovary, and testis. AHR mRNA was detected in all samples tested, and in large amounts in embryos and testis (Fig. 8C).

DISCUSSION

TCDD-induced vascular and bone damages through hyperactivation of AHR

TCDD is the most potent toxicant for vertebrate species. Exposure of vertebrate embryos to TCDD can result in various acute and chronic toxicities such as reproductive failure, teratogenic abnormalities, and immunological dysfunction (Peterson *et al.*, 1993). In fish, vascular damage is the most pronounced adverse effects of TCDD exposure during embryonic development. Vascular hemorrhaging, regression of blood vessels, pericardial sac edema, and reduced circulation are hallmark indicators that vascular function is compromised in the developing embryos (Cantrell *et al.*, 1996; Henry *et al.*, 1997; Hornung *et al.*, 1999; Guiney *et al.*, 2000). The vascular lesions have been demonstrated to be associated with apoptosis and induced expression of Cyt P450 1A in blood vessels of medaka embryos (Cantrell *et al.*, 1998). In the present study, we re-examined the TCDD-induced vascular damage in medaka embryos by observing blood clotting and regression of blood vessels. We found that these vascular damages can be suppressed, but transiently, by antagonist, NF (Fig. 3), giving a convincing evidence that the TCDD-induced vascular damage is mediated through hyperactivation of AHR. The transient suppression may be explained by the fact that TCDD, but not NF, is very stable *in vivo* against catabolic activities of Cyts P450 (Miniero *et al.*, 2001). Although the damage can also be suppressed by Cyts P450 inhibitor, PBO (Fig. 4C), general oxidative stress caused by Cyts P450-mediated oxidative reactions may not be responsible for the

TCDD-induced damage, in inconsistent with the previous conclusion (Cantrell *et al.*, 1996), because reducing agent, NAC, could not recover the damage in vasculature (Fig. 5) or also in bone (data not shown). We assume that a toxic compound that may be accumulated *in vivo* by elevated levels of Cyt P450 is responsible for the TCDD-induced pathology (Fig. 9).

We also found that embryonic treatment with picomolar concentrations of TCDD causes malformation of bone in adult fish (Fig. 7). The treatment did not give apparent complications including blood clotting in the hatching fry, thus, the bone staining is the most sensitive method for detecting TCDD toxicity. The bone deformity could also be recovered by co-treatment with the antagonist (Fig. 7C), implying the role of hyperactivated AHR. TCDD may directly act on bone, because it inhibits osteogenesis in bone-forming cultures of chicken and rat cells (Gierthy *et al.*, 1994; Singh *et al.*, 2000). Treatment of medaka fish with TCDD from the egg stage to post-hatching also caused developmental defects in bone formation at the posterior region of vertebral column and at spines (Figs. 7D, E). However, it may be possible that these defects occurred secondarily to vascular damage, because blood clots formed at the base of the caudal fin under the same condition (Fig. 2Q).

AHR is required for prevention of blood clotting and for proper development of vasculature and bone in medaka fish

AHR is conserved among vertebrates, and ubiquitously expressed in embryos and adult tissues. In the present study, we have cloned three different cDNAs encoding two AHR homologs from medaka fish, *O. latipes* (Fig. 8). The two homologs obtained may belong to a type of AHR1 by amino acid sequence similarity, thus named AHR1 α and AHR1 β . They differ from each other only in C-terminal short peptide, and may be derived from alternative splicing. AHR1 mRNA was also ubiquitously expressed in medaka embryos and adult tissues, suggesting developmental and physiolog-

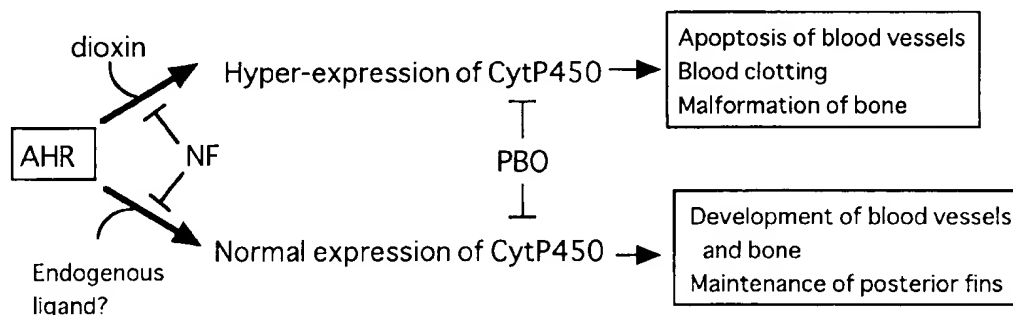


Fig. 9. Model for the role of AHR in the TCDD (dioxin)-induced toxicity, the development of blood vessels and bone, and the maintenance of posterior fins in the medaka fish, *O. latipes*. TCDD-bound AHR induces hyper-expression of a certain Cyt P450, resulting in the toxicities such as apoptosis of blood vessels, blood clotting, and malformation of bone. Either the antagonist (NF) or the Cyts P450 inhibitor (PBO) can suppress the TCDD-induced toxicity. An endogenous ligand is bound to and constitutively activates AHR. The activated AHR is responsible for normal expression of a certain Cyt P450 that is required for the development of blood vessels and bone and homeostasis of posterior fins. *In vivo* inhibition of AHR and Cyt P450 by NF and PBO, respectively, causes developmental abnormalities in vasculature and bone.

ical roles in medaka fish.

To investigate the role of AHR in fish development and physiological homeostasis, medaka embryos (12 hpf) were treated with the antagonists, NF and Res. These compounds did not cause any apparent defects until 4 dpf, but displayed developmental toxicities such as blood clotting and regression of blood vessels at 5 dpf (Figs. 1 and 2). Blood clotting may be caused by regression of blood vessels, because platelet adhesion to subendothelial collagens and activation by components of the extracellular matrix are crucial for blood coagulation (Nieswandt *et al.*, 2001). NF also caused the malformation of bone at 5-day post-hatching (Fig. 7D) and the regression of posterior fins such as anal, caudal, and dorsal fins at the adult period (Fig. 7F). These results suggest the presence of an endogenous ligand for AHR and that constitutive activation of AHR is specifically required for the development of blood vessels and bone and for the maintenance of posterior fins (Fig. 9).

Ligand-bound AHR activates transcription of a battery of genes including various Cys P450. If levels of a certain Cyt P450 were controlled by AHR bound to an endogenous ligand and required for proper development of blood vessels and bone, the well-known inhibitor (PBO) of the enzymatic activity of Cys P450 would induce the same developmental defect as did the antagonist. Treatment of embryos with PBO specifically induced blood clotting, regression of blood vessels (Figs. 1 and 2), and degeneration of the posterior end of spinal cord (data not shown) at the same developmental stage as did the antagonist, suggesting the importance of Cyt P450, the identity of which is, however, unknown (Fig. 9). The synergistic effects exerted by NF and PBO (Fig. 6) also support the hypothesis. We assume that a certain Cyt P450 is responsible for degradation (or catabolism) of a toxic compound that caused the developmental abnormalities.

ACKNOWLEDGEMENTS

We thank S. Omura and R. Horie for assistance and discussion. This study was supported in part by a grant from Iyaku-shigen Research Foundation.

REFERENCES

- Abbott BD, Schmid JE, Pitt JA, Buckalew AR, Wood CR, Held GA, Diliberto JJ (1999) Adverse reproductive outcomes in the transgenic Ah receptor-deficient mouse. *Toxicol Appl Pharmacol* 155: 62–70
- Adams NH, Levi PE, Hodgson E (1993) Regulation of cytochrome P-450 isozymes by methylenedioxyphenyl compounds. *Chem Biol Interact* 86: 255–274
- Cantrell SM, Joy-Schleizinger J, Stegeman JJ, Tillitt DE, Hannink M (1998) Correlation of 2,3,7,8-tetrachlorodibenzo-*p*-dioxin-induced apoptotic cell death in the embryonic vasculature with embryotoxicity. *Toxicol Appl Pharmacol* 148: 24–34
- Cantrell SM, Lutz LH, Tillitt DE, Hannink M (1996) Embryotoxicity of 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD): The embryonic vasculature is a physiological target for TCDD-induced DNA damage and apoptotic cell death in medaka (*Oryzias latipes*). *Toxicol Appl Pharmacol* 141: 23–34
- Casper RF, Quesne M, Rogers IM, Shirota T, Jolivet A, Milgrom E, Savouret JF (1999) Resveratrol has antagonist activity on the aryl hydrocarbon receptor: Implications for prevention of dioxin toxicity. *Mol Pharmacol* 56: 784–790
- Ciolino HP, Daschner PJ, Yeh GC (1998) Resveratrol inhibits transcription of CYP1A1 *in vitro* by preventing activation of the aryl hydrocarbon receptor. *Cancer Res* 58: 5707–5712
- Dahl AR, Hodgson E (1979) The interaction of aliphatic analogs of methylenedioxyphenyl compounds with cytochromes P-450 and P-420. *Chem Biol Interact* 27: 163–175
- Dolwick KM, Schmidt JV, Carver LA, Swanson HI, Bradfield CA (1993) Cloning and expression of a human Ah receptor cDNA. *Mol Pharmacol* 44: 911–917
- Fernandez-Salguero PM, Hilbert DM, Rudikoff S, Ward JM, Gonzalez FJ (1996) Aryl-hydrocarbon receptor-deficient mice are resistant to 2,3,7,8-tetrachlorodibenzo-*p*-dioxin-induced toxicity. *Toxicol Appl Pharmacol* 140: 173–179
- Fernandez-Salguero P, Pineau T, Hilbert DM, McPhail T, Lee SS, Kimura S, Nebert DW, Rudikoff S, Ward JM, Gonzalez FJ (1995) Immune system impairment and hepatic fibrosis in mice lacking the dioxin-binding Ah receptor. *Science* 268: 722–726
- Gasiewicz TA, Rucci G (1991) Alpha-naphthoflavone acts as an antagonist of 2,3,7,8-tetrachlorodibenzo-*p*-dioxin by forming an inactive complex with the Ah receptor. *Mol Pharmacol* 40: 607–612
- Gierthy JF, Silkworth JB, Tassinari M, Stein GS, Lian JB (1994) 2,3,7,8-Tetrachlorodibenzo-*p*-dioxin inhibits differentiation of normal diploid rat osteoblasts *in vitro*. *J Cell Biochem* 54: 231–238
- Gonzalez FJ, Fernandez-Salguero P (1998) The aryl hydrocarbon receptor. *Drug Metab Dispos* 26: 1194–1198
- Guiney PD, Smolowitz RM, Peterson RE, Stegeman JJ (1997) Correlation of 2,3,7,8-tetrachlorodibenzo-*p*-dioxin induction of cytochrome P4501A in vascular endothelium with toxicity in early life stages of lake trout. *Toxicol Appl Pharmacol* 143: 256–273
- Guiney PD, Walker MK, Spitsbergen JM, Peterson RE (2000) Hemodynamic dysfunction and cytochrome P4501A mRNA expression induced by 2,3,7,8-tetrachlorodibenzo-*p*-dioxin during embryonic stages of lake trout development. *Toxicol Appl Pharmacol* 168: 1–14
- Hahn ME, Karchner SI (1995) Evolutionary conservation of the vertebrate Ah (dioxin) receptor: amplification and sequencing of the PAS domain of a teleost Ah receptor cDNA. *Biochem J* 310: 383–387
- Hankinson O (1995) The aryl hydrocarbon receptor complex. *Annu Rev Pharmacol Toxicol* 35: 307–340
- Henry TR, Spitsbergen JM, Hornung MW, Abnet CC, Peterson RE (1997) Early life stage toxicity of 2,3,7,8-tetrachlorodibenzo-*p*-dioxin in zebrafish (*Danio rerio*). *Toxicol Appl Pharmacol* 142: 56–68
- Hornung MW, Spitsbergen JM, Peterson RE (1999) 2,3,7,8-tetrachlorodibenzo-*p*-dioxin alters cardiovascular and craniofacial development and function in sac fry of rainbow trout (*Oncorhynchus mykiss*). *Toxicol Sci* 47: 40–51
- Karchner SI, Powell WH, Hahn ME (1999) Identification and functional characterization of two highly divergent aryl hydrocarbon receptors (AHR1 and AHR2) in the teleost *Fundulus heteroclitus*. *J Biol Chem* 274: 33814–33824
- Kawahara T, Okada H, Yamashita I (2000) Cloning and expression of genomic and complementary DNAs encoding an estrogen receptor in the medaka fish, *Oryzias latipes*. *Zool Sci* 17: 643–649
- Kawahara T, Yamashita I (2000) Estrogen-independent ovary formation in the medaka fish, *Oryzias latipes*. *Zool Sci* 17: 65–68
- Merchant M, Krishnan V, Safe S (1993) Mechanism of action of alpha-naphthoflavone as an Ah receptor antagonist in MCF-7

- human breast cancer cells. *Toxicol Appl Pharmacol* 120: 179–185
- Mimura J, Yamashita K, Nakamura K, Morita M, Takagi TN, Nakao K, Ema M, Sogawa K, Yasuda M, Katsuki M, Fujii-Kuriyama Y (1997) Loss of teratogenic response to 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD) in mice lacking the Ah (dioxin) receptor. *Genes Cells* 2: 645–654
- Miniero R, De Felip E, Ferri F, di Domenico A (2001) An overview of TCDD half-life in mammals and its correlation to body weight. *Chemosphere* 43: 839–844
- Nieswandt B, Brakebusch C, Bergmeier W, Schulte V, Bouvard D, Mokhtari-Nejad R, Lindhout T, Heemskerk JWM, Zirngibl H, Faessler R (2001) Glycoprotein VI but not $\alpha 2\beta 1$ integrin is essential for platelet interaction with collagen. *EMBO J* 20: 2120–2130
- Peterson RE, Theobald HM, Kimmel GL (1993) Developmental and reproductive toxicity of dioxins and related compounds: cross-species comparisons. *Crit Rev Toxicol* 23: 283–335
- Robles R, Morita Y, Mann KK, Perez GI, Yang S, Matikainen T, Sherr DH, Tilly JL (2000) The aryl hydrocarbon receptor, a basic helix-loop-helix transcription factor of the PAS gene family, is required for normal ovarian germ cell dynamics in the mouse. *Endocrinol* 141: 450–453
- Rowlands JC, Gustafsson JA (1997) Aryl hydrocarbon receptor-mediated signal transduction. *Crit Rev Toxicol* 27: 109–134
- Singh SUN, Casper RF, Fritz PC, Sukhu B, Ganss B, Girard Jr B, Savouret JF, Tenenbaum HC (2000) Inhibition of dioxin effects on bone formation *in vitro* by a newly described aryl hydrocarbon receptor antagonist, resveratrol. *J Endocrinol* 167: 183–195
- Takeuchi K (1960) A study of the mutant (*wavy*) in the medaka, *Oryzias latipes*. *Annotationes Zoologicae Japonenses* 33: 124–131
- Testa B, Jenner P (1981) Inhibitors of cytochromes p450 and their mechanisms of action. *Drug Metab Rev* 12: 1–117
- Yamamoto T (1969) Sex differentiation. In "Fish Physiology 3" Ed by WS Hoar, DJ Randall Academic Press, New York, pp 117–175

(Received September 12, 2001 / Accepted December 3, 2001)

TOP, HOME

Tokio Yamamoto In: "Medaka, Biology and Strains" (T. Yamamoto, ed.), Yugakusya Publ. (1975), pp. 17-29.

Systematics and Zoogeography

The killifishes, or Cyprinodontiforms are small fresh and brackish water fishes of worldwide distribution in tropical and temperate latitudes.

Previous classification of the order Cyprinodontes

The classification of the order Cyprinodontes Agassiz (equivalent to Microcyprini Regan) has been worked out by Gill (1865, 1874), Regan (1909, 1911), Hubbs (1924, 1926) and Myers (1931, 1938). The classification followed here is mostly according to Hubbs and Myers and is cited from Kulkarni (1940) who erected a new family Horaichthyidae represented by a remarkable Indian henpecked killifish, *Horaichthys setnai*. However, substituting for the terms Amblyopsoidea and Poecilioidea, the suborders Amblyopsoidei and Cyprinodontoidei are used here, respectively. The subfamily Tomeurinae is removed from the family Poeciliidae to erect a new family Tomeuridae as suggested by Hubbs in his letter to S. L. Hora (India) in 1938. Representative genera are given in parentheses following family names.

Order Cyprinodontes (Microcyprini)

Suborder Amblyopsoidei

Family Amblyopsidae (*Chologaster*, *Amblyopsis*)

Suborder Cyprinodontoidei

Family Cyprinodontidae (*Cyprinodon*, *Fundulus*,

Aplocheilus, *Panchax*, *Oryzias*)

Family Goodeidae (*Goodea*)

Family Poeciliidae (*Poecilia*, *Gambusia*, *Xiphophorus*)

Family Jenynsiidae (*Jenynsia*)

Family Anablepidae (*Anableps*)

Family Tomeuridae (*Tomeurus*)

Family Adrianichthyidae (*Adrianichthys*, *Xenopoecilus*)

Family Phallostethidae (*Phallostethus*, *Gulaphallus*)

Family Horaichthyidae (*Horaichthys*)

The classification listed here has been generally held by ichthyologists until 1962.

As to the status of *Oryzias*, Myers (1931) considered it to represent a monogeneric tribe of the subfamily Fundulinae. Later (1956), he revised his earlier classification, and considered *Oryzias* to represent a monogeneric subfamily of the Cyprinodontidae, the Oryziatinae.

Classification of new order Atherinoformes

Rosen (1962) presented evidence which indicates a relationship of the Amblyopsidae (North American cave fishes) with the percopsiform genera and, more distantly with the gadiforms. He isolated the cave fishes as a new order, the Amblyopsiformes, and recommended its alignment near the Percopsiformes and Gadiformes in a phyletic sequence.

In 1964, Rosen has made drastic taxonomic re-arrangements of the halfbeaks, killifishes, silversides, and their relatives. The outset of his re-arrangements was osteological analyses of the adrianichthyid fishes of Celebes, which were found to have a mixture of beloniform, cyprinodontiform, and mugilform features. Then, his investigation was broadened to include representatives of all these groups as well as a species of phallostethid.

In consequence of reasonable osteological diagnoses, he erected a new order Atherinoformes which includes the excoetoids, scomberesocoids. On the basis of osteological evidence, he separated the medaka (*Oryzias*) from cyprinodontoids, placed it in adrianichthyoids and erected a new family Oryziatidae.

To visualize Rosen's account on osteological difference between cyprinodontoids and adrianichthyoids, the presentation of the schema of the skull of the generalized teleosts as shown in Fig. 2-1 may be apropos.

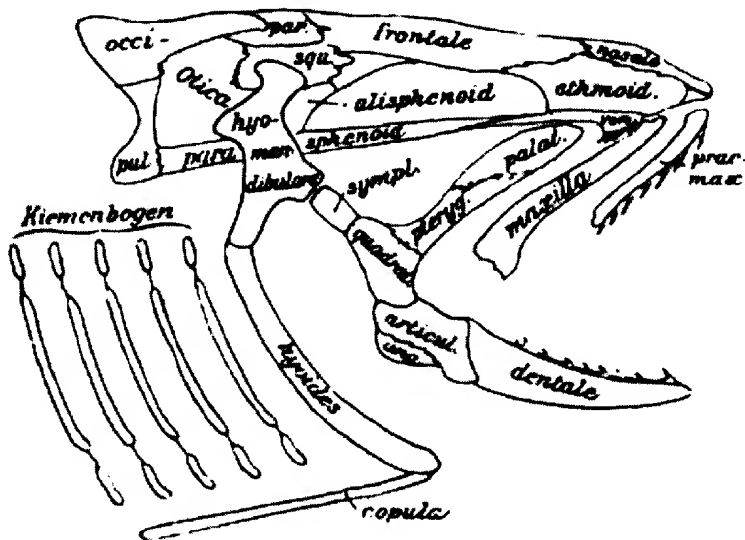


Fig. 2-1. A diagram of teleostean skull. Opercula and Infraorbitalia are removed. ang. = angular, articul. = articular, occi. = occipital, palat. = palatine, p = quadrate, squ. = squamose, sympl. = symplectic, vom. = vomer.

After R. Goldschmidt' E. Selenkas Zoologisches Taschenbuch fur Studierende. 1912 Leipzig, George Thieme.

In cyprinodontoid killifishes, bones of the jaws and the palatoquadrate arch are in such a construction that the premaxilla is protractile. In adrianichthyoid killifishes, on the other hand, the premaxillae are not protractile. The Adrianichthyidae are fishes of small size confined to the fresh-water lakes of Celebes. Two species, *Adrianichthys kruyti* and *Xenopoecilus sarasinorum*, are known. *Xenopoecilus* is characterized by having a large horse-shoe shaped mouth, an enormous ethmoideum and a single, median supraoccipital process formed by fusion of embryologically paired elements; "a cup-like excavation on the distal tip of the autopalatine that is capped by a large ball of cartilage and a discoidal sesamoid bone; a dorsal enlargement of the palatopterygoid arch with a prefrontal (Fig. 2-2); a maxilla that is carried on the upper edge rather than on the outer face of the posterior end of the premaxilla; a premaxilla that lacks a hooked or pointed posteroventral process; a tremendously reduced articular bone without a coronoid process that is almost wholly contained within the posterior part of the dentary; the articulation of the first pleural rib on the third rather than on the second vertebra; pelvic girdles that are not in contact medially and that have a long lateral spur extending upward between ribs; a dorsoventrally asymmetrical caudal skeleton with one or two very slender, rod-like epurals, and a caudal fin that is divided into indistinct upper and lower lobes by having a large gap between rays that articulate with the upper and lower hypural plates on the terminal half-centrum. (Rosen, 1964)

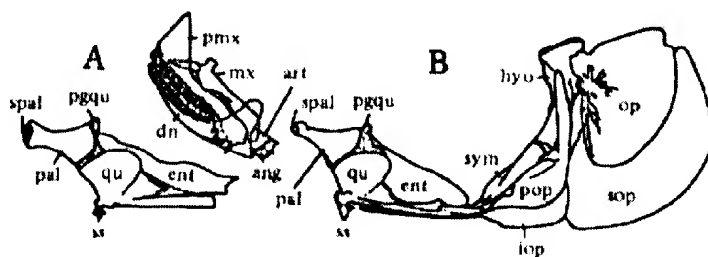


Fig. 2-2. Jaws and jaw suspension in adrianichthyoid killifishes. A. Jaws and palatopterygoid arch in *Oryzias latipes* (Temminck and Schlegel). b. Jaw suspension and opercular apparatus in *Xenopoecillus sarasinorum* (Popta). Note sesamoid bone below quadrate and bony cap over tip of palatine in A. and B. Note in A that lower arm of premaxilla lies over maxilla, large coronoid process on dentary, and absence of similar coronoid elevation on articular. Ang = angular, art = articular, dn = dentary, ent = entpterygoid (mesopterygoid), hyo = hyomandibular, iop = interoperculum, mx = maxilla, op = operculum, pal = palatine (autopalatine with or without dermopalatine), pgqu = pterygoquadrate cartilage, pmx = sesamoid bone capping autopalatine, ss = sesamoid bone, sym = symplectic. Rosen, 1964.

Rosen pointed out that except for the enlarged jaws and the presence of a median supraoccipital process, all the above features described within quotation marks can be identified in *Oryzias* (Fig. 2-2) but in no other killifishes so far as known.

It is therefore apparent that adrianichthyids and the medaka are intimately related and that they constitute a distinct subgroup of the killifishes, the adrianichthyoids, containing the families Adrianichthyidae (*Adrianichthys* and *Xenopoecilus*), Oryziatidae (*Oryzias*), and Horaichthyidae (*Horaichthys*), in contrast to the remainder of the families which are grouped together as cyprinodontoids (Cyprinodontoidea). Basing on Rosen's (1964) findings, Turner (1965) conveniently enumerated difference between cyprinodontoids and *Oryzias* as follows:

Cyprinodontoidae	<i>Oryzias</i>
1. First pleural rib on second vertebra.	First pleural rib on third vertebra.
2. Pelvic girdle bones joined mid-ventrally; no upright lateral spur.	Pelvic girdle bones not joined mid-ventrally; an upright lateral spur present.
3. Lower end of premaxilla bone expanded or hooked and sandwiched between the lower end of maxilla bone and dentary bone (lower jaw).	Lower end of premaxilla bone not expanded, and dorsal to the maxilla bone rather than between it and the dentary bone.
4. Hypural plates often fused.	Hypural plates never fused.
5. Hypochordal musculature entirely absent.	Hypochordal musculature present.
6. Caudal fin never incipiently lobed.	Caudal fin incipiently lobed.

The family Horaichthyidae erected by Kulkarni (1940) comprises a single species, *Horaichthys setnai*. It is a small translucent oviparous fish inhabiting brackish waters and estuaries in the province of Bombay, India. Osteological study (Kulkarni 1948) showed that its head skeleton is closely allied to that of *Oryzias* but greatly different from that of *Aplocheilichthys*. *Horaichthys*, however, is different from known species of *Oryzias* in having a larger number of the anal fin- rays (about 28 to 32).

In the male, six anterior rays of the anal fin are separated from the rest of the fin and modified into an elaborate male organ (gonopodium). Of six rays the third, fourth and fifth ones are profoundly modified forming the 3- 4-5 complex. (Fig. 2-3). In the female right pelvic fin is usually absent. The genital opening of the female is situated on the left ventral side and is surrounded by genital pads. *Horaichthys* is supposed to have evolved from *Oryzias*, but as the development of the gonopodium in association with the henpecked sexual behavior is so remarkable that Kulkarni (1940) has proposed to erect a new family rank for this fish.

The male appears to be always afraid of the female which on occasions chases him away. At the time of mating, "the male swims below and behind her at a distance of about 2 to 3 cm. He then darts towards her on the left with almost lightning speed. As he approaches his mate he lashes out the gonopodium sideways almost at right angles to his body and strikes its terminal end against her genital opening. The spermatophores are transferred to the female in this momentary contact, and become attached by their distal hooks."

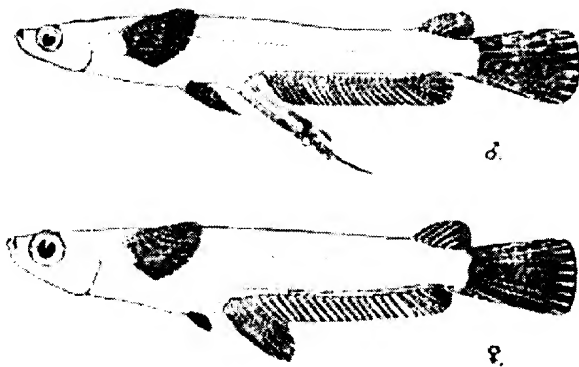


Fig.2-3. Lateral view of a male and a female specimen of *Horaichthys setnai*.
x 4 Kulkalni, 1940.

A special feature of *Horaichthys* is that the testis produces special sperm capsules of spermatophores (2-300 in number) instead of ordinary semi-fluid milt with suspended sperms.

A spermatophore is a tiny hyaline body (0.6 mm long and 0.1 mm thick), the broad part of which contains mass of sperms. At the tapering end, there is a pointed cap with stiff hooks and barb-like structures which point backwards. It is with the aid of these hooks and barbs that the spermatophore get attached near the genital opening of the female.

There is no permanent opening on the spermatophore for the liberation of sperms. Before liberation of sperms, a small bulging appears at the neck of the tapering spermatophore and begins to enlarge. When the protuberance becomes sufficiently large, an opening is formed at its tip by rupture of membrane and sperms are liberated. They swim into the genital pore of the female.

The following is the classification of the new order Atheriniiformes by Rosen (1964), representative species being given in parentheses.

Suborder Exocoetoidei

Superfamily Exocoetoidea

Family Hemiramphidae (*Hemiramphus*)

Family Exocoetidae (*Exocoetus*)

Superfamily Scomberesocoidea

Family Belonidae (*Ablennus*)

Family Scomberesocidae (*Cololabis*)

Suborder Cyprinodontoides

Superfamily Adrianichthyoidea

Family Oryziatidae (*Oryzias*)

Family Adrianichthyidae (*Adrianichthys*, *Xenopoecilus*)

Family Horaichthyidae (*Horaichthys*)

Superfamily Cyprinodontoides

Family Cyprinodontidae (*Fundulus*, *Aplocheilus*)
Family Goodeidae (*Goodea*)
Family Jenynsiidae (*Jenynsia*)
Family Anablepidae (*Anableps*)
Family Poeciliidae (*Poecilia*, *Xiphophorus*)
Suborder Atherinoidei
 Superfamily Atherinoidea
 Family Melanotaeniidae
 Family Atherinidae (*Atherina*)
 Family Isonidae, new family (*Iso*)
 Superfamily Phallostethoidea
 Family Neostethidae (*Neostethus*)
 Family Phallostethidae (*Phallostethus*)

The family Oryziatidae

Rosen (1964) erected a new monogeneric family and described the following diagnoses of the family Oryziatidae. Type genus: *Oryzias* Jordan and Snyder, 1906. Diagnoses: The Oryziatidae differ from their closest relatives, the adrianichthyids, in lacking the tremendously enlarged jaws and ethmoideum, in having paired supraoccipital processes (rather than a single median process), and in having the inferior pharyngeal bone distinctly separated (rather than united), and from all cyprinodontoids as follows: autopalatine usually capped by sesamoid bone; pterygoquadrate cartilage forming dorsal process; lower end of premaxilla not hooked or trapezoidal, situated below maxilla rather than between maxilla and dentary bone; first pleural rib on third vertebra; supracleithrum wanting; pelvic bones with upright lateral spurs and not joined midventrally; hypochordal musculature present on caudal fin.

Composition: Rosen listed following seven species of a single genus, *Oryzias*: *O. latipes* (Temminck and Schlegel), *O. melastigma* (McClelland), *O. celebensis* (Weber), *O. timorensis* (Weber and de Beaufort), *O. javanicus* (Bleeker), *O. curvinotus* (Nichols and Pope), and *O. minutillus* Smith. To these, *O. luzonensis* (Herre and Ablan) may be added. Besides these, Turner (1965) mentioned *O. matenensis* (Aurich), and *O. marmoratus* (Aurich) from the Celebes.

Probably not all these nominal species are valid, since some nominal species are different only in the anal fin-ray frequency.

The genus *Oryzias*

The following is the diagnoses of the genus *Oryzias* described by Jordan and Snyder (1906), basing on *O. latipes* which has previously been known as *Aplocheilus latipes*.

Body elliptical in form, compressed, covered with large scales; mouth small, with two rows of small, simple, pointed teeth; *no teeth on vomer**1; gill-opening not restricted above; intestinal canal short, about as large as body; peritoneum black. Dorsal fin short, inserted above middle of anal; anal very long seventeen to twenty rays; caudal fin truncate. *Sexes similar**2 *except color*; anal fin not modified in the male. *1 Kulkarni(1948) first showed that *os vomer* is absent in *Oryzias melastigma*. *2 Sexual

dimorphism is prominent. See Chap. 8.

The species *Oryzias latipes*

The following description by Oshima (1919) based on a specimen of *Oryzias latipes* collected from Shori, Formosa is cited here as the diagnoses of the species since it is very precise and correct excepting two words starred and daggered.

Head 4 in length (body length divided by head length is 4); depth 4.5; depth of caudal peduncle 9.5; eye 2.5 in head (head length divided by eye diameter is 2.5); interorbital space 2; snout 4; D.6; A.18; P.9; V.5; thirty one scales in a lateral series; five branchiostegals.

Posterior half of the body compressed, becoming broader anteriorly, highest in front of the anal; head flattened; interorbital space broad; snout shorter than the diameter of eye, broadly rounded anteriorly; mouth anterior, transverse; lower jaw slightly projecting, each jaw with two rows of minute pointed teeth, those on the posterior row smaller; vomer*1 smooth; thirteen short, pointed gill-rakers on the first arch; eyes very large, anterior and superior.

Dorsal fin short, on the posterior half of body, its origin above the posterior two thirds of anal, its height equal to the distance between tip of snout and posterior margin of orbit; pectoral inserted on the median line of body; the ventral small, reaching vent; base of the anal very long, its posterior end opposite to that of the dorsal, anterior ray longest; tip of the caudal fin *rounded*.*2

Top and sides of head, throat, and chin naked; body covered with cycloid scales, lateral line absent.

Color in formalin pale gray above, lower parts silvery; a black longitudinal streak from the nape to the origin the dorsal; sides of body with a faint dusky stripe along the middle line, top of head dark; the edges of scales dusky; fin-rays of the ventral and anal dotted with minute black spots; all the fins whitish; peritoneum black. Length of body 28 mm.

Habitat: The present species is very common in rice-fields and pools on the island.

*1 Vomer is absent in *Oryzias* in reality.

*2 The caudal fin is almost truncate, strictly, however, it is incipiently lobed.

Change of nomenclature of the medaka

The medaka was first described as *Poecilia latipes* by Temminck and Schlegel in 1846 (Siebold's Fauna Japonica, Poiss., P.224, Pl.102, Fig. 5). Gŷnther changed it as *Haplochilus latipes*. Jordan and Snyder (1901) described it as *Aplocheilus latipes* but later they separated it from *Aplocheilus* and erected a new genus *Oryzias*. They regarded *Oryzias* as having no teeth on vomer*1 while *Aplocheilus* possesses teeth on it.

Myers (1931) placed the medaka in the tribe Aplocheilini of the subfamily Fundulinae in the family Cyprinodontidae. He stated that the chief character of fishes of the tribe is the non-protractile premaxillae. The pectoral fin are set high and pseudobranchiae and vomerine teeth are never present. The species range from Japan and Central China south to Celebes and Timor and west to Southern India. A single genus, *Aplocheilus*, of which *Oryzias* is a synonym*2. Smith, (1945) pointed out that the genus known as Panchax is a synonym of *Aplocheilus* McClelland and *Aplocheilus* Weber and de Beaufort is a synonym of *Oryzias* Jordan and Snyder. He described *Aplocheilus panchax* (Hamilton) and *Oryzias minutillus* n. sp. from Thailand.

According to him, the two genera may be distinguished by the following characters:

- a. Upper jaw protractile; mouth moderate size with its corners abruptly bent downward; vomer toothed; pseudobranchiae present; branchial membranes free from each other and from isthmus; pectoral fins with their upper base at or below longitudinal axis of body *Aploc*
- b. Upper jaw not protractile; mouth small with its corners obtusely bent downward; vomer toothless; no pseudobranchiae; branchial membranes united across isthmus; pectoral fins with upper base well above longitudinal axis of body *Oryzia*

The correct scientific name of the medaka is *Oryzias latipes* (Temminck and Schlegel).

From Jordan and Snyder (1906) onwards, all taxonomists stated that *Oryzias* has toothless vomer while *Aplocheilus* has toothed vomer. Kulkarni (1948) has made a precise osteological study of Indian killifishes and found that vomer is absent in both *Oryzias melastigma* and *Horaichthys setnai* while *Aplocheilus lineata* possesses toothed vomer.

*1 Vomer is absent in *Oryzias* in reality.

*2 Now, the two genera belong to the different super-families.

Geographical distribution of species belonging to the Genus *Oryzias*

All the species of the genus *Oryzias* are distributed in India, South Asia, the Indo-Australian archipelago and the Far East. Their habitats are widely ranged from tropical, subtropical, and temperate regions as shown in Figure 2-4 and in the following lines.

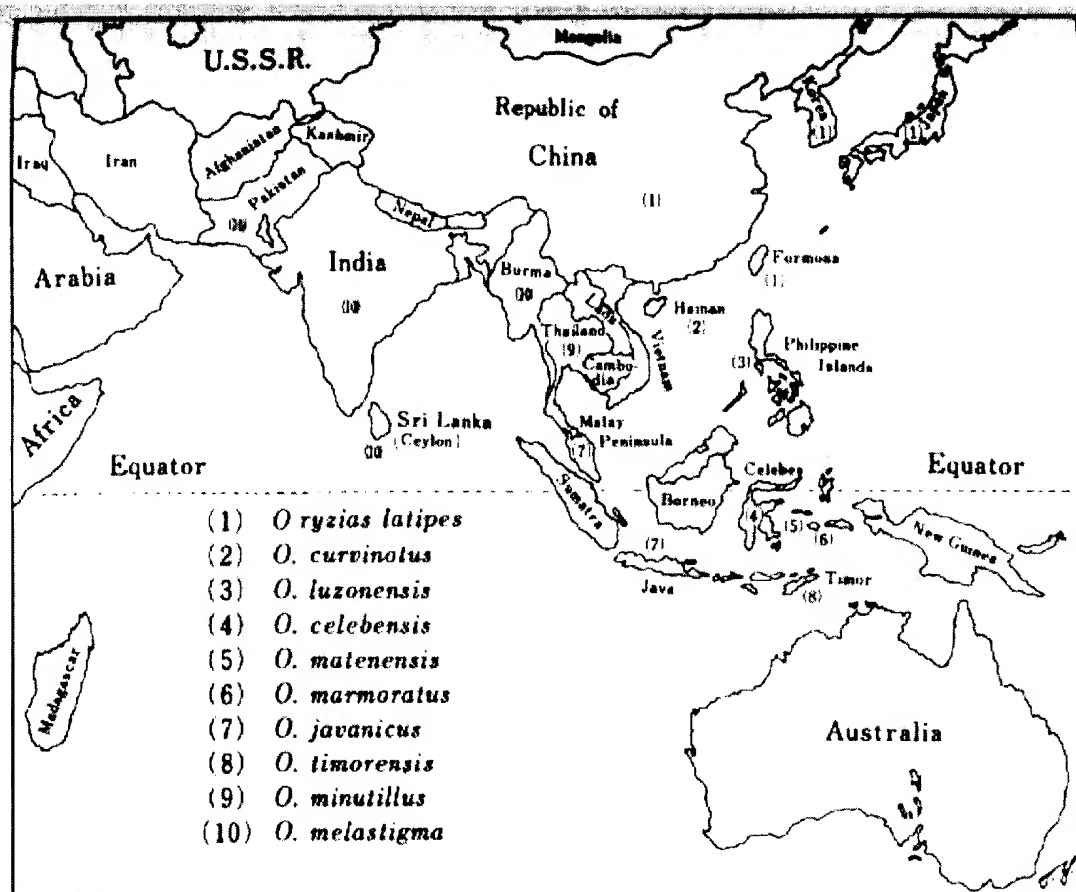


Fig. 2-4. A

zoogeographical map showing distribution of species of the Genus *Oryzias*. Original.

- (1) *O. latipes* (Temminck and Shlegel): Japan, Korea, Formosa, and China
- (2) *O. curvinotus* (Nicols and Pope): The island of Hainan.
- (3) *O. luzonensis* (Herre and Ablan): Luzon in the Philippines.
- (4) *O. celebensis* (Weber): The Celebes.
- (5) *O. matenensis* (Aurich): The Celebes.
- (6) *O. marmoratus* (Aurich): The Celebes.
- (7) *O. javanicus* (Bleeker): The Indo-Malaysian archipelago and Malaya.
- (8) *O. timorensis* (Weber and de Beaufort): The island of Timor.
- (9) *O. minutillus* Smith: Thailand.
- (10) *O. melastigma* (McClelland): India, Western Pakistan, and Sri Lanka (Ceylon).

In the main, all the *Oryzias* species are fresh-water fishes. *O. latipes* and *O. melastigma* inhabit both fresh and brackish water. *O. latipes* is so tolerate slinity that it thrives in tide pools in Korea and Kyushu in Japan.

References

- Gill, T.N., 1965 Synopsis of the fishes in the Gulf of St. Lawrence and Bay of Fundy. Canadian Nat., Ser. 2, 2: (No.4) 244-266.
- Gill, T.N., 1874 Arrangement of the families of fishes, or Classes Pisces, Marssipobranchii and Leptocardii. Smithonian Misc. Coll., for 1872, 11: (No.247) 1-49.

- Herre, A.W., and G.L. Ablan, 1934 *Aplocheilus luzonensis*, a new Philippine Cyprinodont. Philippine Jour. Sci., 54: (No.2) 275-277.
- Hubbs, C.L., 1924 Studies on the fishes of the order Cyprinodontes I.-IV. Misc. Publ. Mus. Zool., Univ. Michigan, No. 13: 1-31.
- Hubbs, C.L., 1926 Studies on the fishes of the order Cyprinodontes VI. Misc. Publ. Mus. Zool., Univ. Michigan, No. 16: 1-87.
- Jordan, D.S., and J.O. Snyder, 1906 A review of the Poeciliidae or killifishes of Japan. Proc. U.S. Nat. Mus., 31: 287-290.
- Kulkarni, C.V., 1940 On the systematic position, structural modifications, bionomics and development of a remarkable new family of cyprinodont fishes from the province of Bombay. Rec. Indian Mus., 42: 379-423.
- Kurkarni, C.V., 1948 The osteology of Indian cyprinodonts. Part. I. comparative study of the head skeleton of *Aplocheilus*, *Oryzias* and *Horaichthys*. Proc. Natl. Inst. Sci. India, 14: (No.2) 65-119.
- McClelland, J., 1839 Asiatic researches, 19: 301.
- Myers, G.S., 1931 The primary groups of oviparous cyprinodont fishes. Stanford Univ. Publ. Biol. Sci. VI. No. 3: 7-14.
- Myers, G.S., 1938 Studies on the genera of cyprinodont fishes. Copeia (1938): 136-143.
- Nichols, J.T., and C.H. Pope, 1927 The fishes of Hainan. Bull. Amer. Mus. Nat. Hist., 54: 321.
- Oshima, M., 1919 Contributions to the study of the fresh water fishes of the island of Formosa. Annals Carnegie Mus., 12: 169-328.
- Regan, C.T., 1909 The classification of teleostean fishes. Ann. Mag. Nat. Hist., Ser. 8, 3: 75-86.
- Regan, C.T., 1911 The osteology and classification of the teleostean fishes of the order Microcyprini. Ann. Mag. Nat. Hist., Ser. 8, 7: 320-327.
- Rosen, D.E., 1962 Comments on the relationships of the North American cave fishes of the family Amblyopsidae. Amer. Mus. Novitates, No. 2109: 1-35.
- Rosen, D.E., 1964 The relationships and taxonomic position of the halfbeaks, killifishes, silversides, and their relatives. Bull. Amer. Mus. Nat. Hist., 127: (Art.5) 217-268.
- Smith, H.M., 1938 Status of the oriental fish genera *Aplocheilus* and *Panchax*. Proc. Biol. Soc. Washington, 51: 165-166.
- Smith, H.M., 1945 The fresh-water fishes of Siam, or Thailand. U.S. Natl. Mus. Bull., 188: 1-622.
- Turner, B.J., 1965 A new place for the medakas. Classica, 1: (No.2) 1-6.
- Weber, M., 1913 Neue Beitrage zur Kenntnis der Süßwasserfische von Celebes. Bijdr. Dierk., Amsterdam, 19: 197-213.
- Weber, M., and L.F. de Beaufort 1922 The fishes of the Indo-Australian archipelago IV. Heteromi, Solenichthyes, Synentognathi, Percosoces, Labyrinthici, Microcyprini. Leiden, E.J. Brill, Ltd.